

1 **Prevalence of feline lungworm *Aelurostrongylus abstrusus* in England**

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26 **Abstract**

27 Infection of cats with lungworm *Aelurostrongylus abstrusus* has recently been documented in
28 the UK. Here, we aimed to study the prevalence of *A. abstrusus* in fecal samples from cats
29 from England. A total of 950 fecal samples were collected from cats of various ages, breeds,
30 genders, and geographic regions across England. A total of 17 (1.7%) cats were positive for *A.*
31 *abstrusus* based on species-specific morphological features of the larvae isolated by Baermann
32 technique. There was no statistically significant difference in the proportion of positive samples
33 between male (444; 46.7%) and female (506; 53.2%). Multiple regression analysis showed that
34 the prevalence of feline lungworm was significantly different in different geographic regions:
35 in comparison with East Midlands, some regions had shown significantly increased odds of *A.*
36 *abstrusus*-positive samples (South East [odds ratio [OR] = 7.68; 95% confidence interval [CI]
37 = 1.70 to 32.76; $P = 0.01$]; West Midlands [OR = 6.20; 95% CI = 1.21 to 26.84; $P = 0.02$]),
38 while the other regions also had increased odds although not statistically significant (Greater
39 London [OR = 9.63; 95% CI = 0.43 to 84.05; $P = 0.07$]; North West [OR = 4.25; 95% CI =
40 0.59 to 20.89; $P = 0.09$]; South West [OR = 2.48; 95% CI = 0.12 to 17.64; $P = 0.43$]; and North
41 East [OR = 1.88; 95% CI = 0.10 to 12.24; $P = 0.57$]). Keeping cats inside was protective
42 against the risk of infection compared with those having outdoor access (OR = 0.09; 95% CI
43 = 0.01 to 0.48; $P = 0.02$). On the other hand, sex, age, breed, and deworming history did not
44 have any significant effect on the likelihood of infection. Our data indicate that *A. abstrusus* is
45 a parasite of potential significance in cats, in particular those from certain geographic regions.
46 To reduce the spread of this parasite, an integrated feline lungworm control program needs to
47 be implemented.

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49 **Keywords:** Cats; lungworm; *Aelurostrongylus abstrusus*; prevalence; survey; risk factors

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51 1. Introduction

52 The gastropod-borne nematode *Aelurostrongylus abstrusus* (Railliet, 1898) is the most
53 common lungworm of domestic and wild felids and is found in many parts of the world,
54 including Europe, USA, South America and Australia (Scott, 1973; Elsheikha et al., 2016;
55 Giannelli et al., 2017; Penagos-Tabares et al., 2018). This parasite has a considerable impact
56 on cat's health and welfare. Also, it has shown both regional endemicity and geographic
57 expansion across Europe. Infected cats exhibit chronic wasting, cough, dyspnea, pulmonary
58 wheezes and other signs of lower airway disease, although asymptomatic cases, shedding high
59 number of larvae in feces, may also occur (Genchi et al., 2014; Elsheikha et al., 2016; Hansen
60 et al., 2017). In addition to *A. abstrusus*, recent studies have detected other metastrongyloids,
61 such as *Troglostrongylus brevior* (Crenosomatidae) and *Oslerus rostratus* (Filaroididae) and
62 the trichurid *Eucoleus aerophilus* (syn. *Capillaria aerophila*) in the lungs of cats (Pennisi et
63 al., 2015; Giannelli et al., 2017).

64 Some biological and epidemiological drivers (Traversa et al., 2009; Beugnet et al., 2014;
65 Hansen et al., 2017), some of them yet unknown, appear to be increasing the risk of infection
66 in cats in certain parts of the world. However, important gaps remain in the available literature
67 surrounding the prevalence of feline lungworm infection and its epidemiological patterns as
68 well as determinants. Lack of understanding of these changing patterns have serious
69 implications from a clinical standpoint, given that a delay in diagnosis and treatment can lead
70 to severe lesions and even death of the infected cat. Recently, more cases have begun to be
71 observed by clinicians (Gunn-Moore and Elsheikha, 2018). Despite this increasing frequency
72 of *A. abstrusus* in cats in the UK, there is lack of epidemiological studies that assess the
73 prevalence and distribution of this parasite in cats in The UK. A pan-European study involving
74 12 countries, reported 0% *A. abstrusus* infection rate in fecal samples collected from
75 Cambridge, UK (Giannelli et al., 2017). However, this study cannot be representative to the

76 status of *A. abstrusus* infection in The UK due to its very small sample size. Given the paucity
77 of data on *A. abstrusus* in The UK, a larger survey involving more samples collected from
78 diverse geographic areas is needed in order to provide important insight into the transmission
79 potential of *A. abstrusus* in cats The UK. Based on these data, pet owner awareness and
80 education campaigns, launched by pharma and European Scientific Counsel Companion
81 Animal Parasites (ESCCAP), that promote adherence to lungworm prophylaxis can be tailored
82 to at-risk cat populations.

83 We previously conducted a cross-sectional survey in England and in the initial phase we
84 detected *A. abstrusus* larvae in the feces of 2.2% (14 out of 629) of cats (Elsheikha et al., 2017).
85 Herein, we report a more up-to-date *A. abstrusus* prevalence rate, after the completion of the
86 survey, based on the analysis of 950 fecal samples from cats across seven main geographic
87 regions of England. Our study established a new background prevalence of *A. abstrusus* in cats
88 in England and identified outdoor access as a potential risk factor for *A. abstrusus* infection.
89 This new knowledge may lead to more insight into the real burden and risk of feline lungworm
90 infection in the UK, which will ultimately lead to improved sustainable management strategies
91 for feline aelurostrongylosis.

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94 **2. Materials and methods**

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96 *2.1. Fecal samples and data collection*

97 From January 2016 to January 2018, fecal samples ($n = 950$) were collected from cats, 506
98 females and 444 males, across seven administrative regions of England. The study was
99 designed to include feral and street cats in addition to domestic cats. Fecal samples were
100 collected from cats from shelters, catteries and privately owned cats, and were examined using

101 Baermann technique in order to isolate the first stage larvae (L1s). Morphological identification
102 of the isolated *A. abstrusus* L1s and its differentiation from L1s of other metastrongyloids was
103 achieved via microscopic examination using previously described morphometric features of *A.*
104 *abstrusus* larvae (Gerichter, 1949; Brianti et al., 2014; Giannelli et al., 2014; Giannelli et al.,
105 20117).

106 Data on sex (male vs female), age (kitten [0 – 6 months]; junior [7 months – 2 years]; prime
107 [3 years – 6 years]; mature [7 years – 10 years]; and senior [11 years – 14 years]), breed, main
108 geographic regions in England (North East, North west, South East, South West, East
109 Midlands, West Midlands, and Greater London), deworming history (recently treated using
110 anthelmintics, such as emodepside (Profender[®]) or macrocyclic lactones, which have reported
111 efficacy against *A. abstrusus* vs left un-treated), and lifestyle (indoor, outdoor access, feral,
112 and stray) were collected. Ethical approval was granted by the Research Ethics Committee of
113 School of Veterinary Medicine and Science, University of Nottingham.

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115 2.2. Prevalence and risk factor analyses

116 Statistical relationships were assessed between fecal shedding of *A. abstrusus* larvae and
117 defined risk factors, such as age, sex, geographic location, cat lifestyle, and animal treatment
118 status at the time of fecal sampling. The overall parasite prevalence was determined by dividing
119 the number of parasite-positive fecal samples by the total number of samples collected within
120 each risk factor category. Test of independence for contingency tables was used to evaluate
121 associations between each risk factor (e.g., animal sex, age, breed etc.) and presence of *A.*
122 *abstrusus* larvae. Multiple logistic regression was used to investigate the associations between
123 host-specific, demographic, and environmental risk factors with respect to test outcome (e.g.,
124 parasite present or absent). Risk factors that were significant at a *p* level of <0.1 were then
125 incorporated in a forward-stepping manner into multiple logistic regression models. These

126 multivariable models yielded adjusted odds ratios that simultaneously measured the strength
127 of associations between multiple risk factors and the presence of parasite larvae in cat's feces.

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129 **2. Results and discussion**

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131 *2.1. Characteristics of the cat population*

132 A total of 950 cats were examined with ages ranging from 2 to 240 months (mean age 53.1
133 \pm 37.7 months). Approximately, 53.2% (506) of the cats were females and 46.7% (444) were
134 males. Breed distribution included 910 (95.78%) domestic short hair, 27 (2.84%) domestic
135 longhair and 13 (1.36%) belonged to British Semi longhair ($n = 6$), British longhair ($n = 3$),
136 Bengal ($n = 1$), Burmese ($n = 1$), Cornish Rex ($n = 1$), and Maine Coon ($n = 1$).

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138 *2.2. Prevalence of and risk factors associated with infection*

139 Overall, 1.7% (17/950) of the fecal samples tested were positive. This prevalence seems
140 to fall within the range reported in Europe, where prevalence rates varied greatly among
141 European countries, the lowest prevalence was 0.38% recorded in Croatia ([Grabarević et al.,
142 1999](#)) and the highest was 43.1% in Albania ([Knaus et al., 2011](#)). Prevalence of *A. abstrusus*
143 in other European countries ranged from 1% in Germany ([Mundhenke and Dausgchies, 1999](#))
144 to 2.08% in Ireland ([Garcia-Campos et al., 2018](#)), 2.3% in Switzerland ([Zottler et al., 2019](#)),
145 2.6% in the Netherlands ([Robben et al., 2004](#)), 8.3% in the Denmark ([Hansen et al., 2017](#)) to
146 26.5% in in Italy ([Genchi et al., 2014](#)). The prevalence rate of *A. abstrusus* can vary
147 significantly within the same country, for example in Denmark local prevalence rates varied
148 from 0% [95% CI: 0.0–8.8] to 31.4% [95% CI: 16.9–49.3] ([Hansen et al., 2017](#)). Difference in
149 prevalence rates was also detected among three Italian regions ([Giannelli et al., 2017](#)). A
150 similar trend was detected in our study where significant differences were observed in the

151 geographic regions in regard to their association with the increase in the odds of *A. abstrusus*
152 infection, in comparison with East Midlands region (Table 1). The disparity among the
153 prevalences of feline lungworm *A. abstrusus* in certain geographic regions may reflect the level
154 of transmission or availability of intermediate and paratenic reservoir hosts that are able to
155 maintain *A. abstrusus* life cycle in certain areas

156 According to a recent epidemiological survey conducted across 12 European countries,
157 feline lungworms were the second most frequent group of nematodes diagnosed in cats, and
158 although *A. abstrusus* was the most frequently detected lungworm species across Europe, none
159 of the samples tested from cats in Switzerland or Cambridge (UK) was positive for *A. abstrusus*
160 (Giannelli et al., 2017). However, results reported from Cambridge may not be representative
161 of the cat population in the UK due to the very small sample size examined. The broad
162 geographic distribution of *A. abstrusus* in our study indicates that *A. abstrusus* is circulating in
163 cat population and not restricted to a certain locality in England. Further epidemiological
164 studies are required to determine the factors that drive the transmission cycle of *A. abstrusus*
165 in the areas where this parasite is highly prevalent.

166 In this study, the larvae per gram of feces (LPG) were determined using Baermann's
167 technique and ranged from 8 to 22 (11.6 ± 3.3). This was surprisingly low compared to a
168 previous study that detected a mean of 508.7 LPG (Giannelli et al., 2017). The larval survival
169 tend to decline, due to dehydration, depending on the cat litter type and the duration of time
170 faecal samples remain in the litter; a reduction in the viability of 80% of larvae occurred after
171 3 h and reached almost 100% after 24 h (Abbate et al., 2018). In our study, the length of time
172 samples remained in the litter before collection ranged from 1 to 4 h. Thus, we must be
173 cognizant of the potential influence of dehydration on the larval viability, given the low-
174 parasitic load and the time elapsed while the samples are present in the cat litter, which may
175 have underestimated the isolation rate of larvae in our study. It is also worth mentioning that

176 although Baermann's technique is specifically used for direct isolation of lungworm larvae
177 from feces its diagnostic performance, in particular, sensitivity can be compromised by various
178 factors. These include the inability to isolate larvae in the pre-patent period, inconsistent
179 shedding of the larvae especially in cases with low parasite burdens, or cessation of shedding
180 larvae by some cats, despite being infected, which in turn lead to false negative results
181 (Hamilton, 1968; Elsheikha et al., 2016). To increase the accuracy of detection of *A. abstrusus*
182 in future surveys, Baermann's technique should be performed on freshly voided fecal samples
183 collected on three consecutive days. Serological detection of antibodies has dramatically
184 improved the sensitivity of detection of lungworms (Zottler et al., 2017), adding more value
185 for the diagnosis of feline aelurostrongylosis. Thus, a greater emphasis on the use of a
186 serological assay in conjunction with fecal analysis may be warranted to achieve more accurate
187 laboratory diagnosis.

188 We examined the association between age, gender, breed, deworming history and lifestyle,
189 and the risk of infection with *A. abstrusus*. This risk assessment analysis revealed that age, sex,
190 and breed are not significant risk factors of infection. Also, we did not detect any differences
191 between neutered and intact cats. In the present and previous studies, sex was not a risk factor
192 for infection with *A. abstrusus* (Traversa et al., 2008; Barutzki and Schaper, 2013; Olsen et al.,
193 2015; Hansen et al., 2017). Although the risk of *A. abstrusus* infection in Denmark was lower
194 in kittens younger than 11 weeks compared to older cats (Hansen et al., 2017). Another study
195 reported significantly higher prevalence in cats younger than 2 years and in cats co-infected
196 with other gastrointestinal parasites (Giannelli et al., 2017). Our analysis did not detect any
197 effect of the age on the frequency of infection.

198 Out of the 17 infected cats, 13 had outdoor access, three were stray cats and one was an
199 indoor cat (Table 2). Keeping cats indoor was associated with significant protection against
200 infection (odds ratio [OR] = 0.09; 95% confidence interval [CI] = 0.01 to 0.48; $p = 0.02$). In

201 contrast, no significant difference was detected between stray cats and cats with outdoor access
202 (OR = 1.01; 95% CI = 0.22 to 3.49; $p = 0.99$); probably due to the small number of cats in
203 these categories. These findings lend further support to previously reported findings where
204 rural origin, feral lifestyle and outdoor access have been shown to correlate with an increased
205 risk of infection (Traversa et al., 2009; Beugnet et al., 2014; Hansen et al., 2017).

206 Out of the 17 infected cats, four cats were treated with various anthelmintics on the day of
207 sample collection and 13 cats were non-treated. Our analysis has shown a lack of correlation
208 between deworming history and risk of infection. Although statistical analysis did not identify
209 deworming as significantly reducing infection risk, treatment is still likely a key factor that
210 influences the frequency of infection with lungworm in cats. This is because many cats with
211 outdoor access have more opportunity to acquire infection by preying on intermediate and
212 transport hosts. Also, the frequency and timing of deworming are likely to affect the likelihood
213 of infection. Additionally, these results should be interpreted with caution given the small
214 number of positive samples in the stratified categories, which may not have been sufficient to
215 identify any protective effect of deworming.

216 In conclusion, the present study addressed a significant aspect of the epidemiology of *A.*
217 *abstrusus*, a potential serious health problem in feline medicine. Our findings demonstrate that
218 *A. abstrusus* is present in 1.7% of cats in England, and infection frequency seems to vary
219 according to the geographic regions and lifestyle. These findings suggest that the lungworm *A.*
220 *abstrusus* should be considered a potential cause of respiratory tract disease in cats presenting
221 with pulmonary manifestations including cats with mild respiratory signs. However, it is
222 possible that cats can be infected and shed high number of larvae in feces without presenting
223 obvious clinical signs. Therefore, integrated strategies for the effective management of *A.*
224 *abstrusus* as well as other feline metastrongyloid lungworms (*Troglostrongylus* spp., *Oslerus*
225 *rostratus*, *Capillaria aerophila*), should be implemented and can be achieved through using

226 effective preventative anthelmintics, enhanced diagnostics and increased awareness of
227 veterinary professionals of feline lungworms.

228

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235 **References**

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- 237 Abbate, J.M., Arfuso, F., Gaglio, G., Napoli, E., Cavaleria, M.A., Giannetto, S., Otranto, D.,
238 Brianti, E., 2018. Larval survival of *Aelurostrongylus abstrusus* lungworm in cat litters. J
239 Feline Med Surg. Nov 12:1098612X18811168. doi: 10.1177/1098612X18811168. [Epub
240 ahead of print]
- 241 Barutzki, D., Schaper, R., 2013. Occurrence and regional distribution of *Aelurostrongylus*
242 *abstrusus* in cats in Germany. Parasitol. Res. 112, 855-861.
- 243 Beugnet, F., Chalvet-Monfray, K., Cozma, V., Farkas, R., Guillot, J., Halos, L., Joachim, A.,
244 Losson, B., Miro, G., Otranto, D., Renaud, M., Rinaldi, L., 2014. Parasites of domestic
245 owned cats in Europe: co-infestations and risk factors. Parasit. Vectors 7,: 291.
- 246 Brianti, E., Giannetto, S., Dantas-Torres, F., Otranto, D., 2014. Lungworms of the genus
247 *Troglostrongylus* (Strongylida: Crenosomatidae): neglected parasites for domestic cats.
248 Vet. Parasitol. 202, 104-112.

249 Elsheikha, H.M., Schnyder, M., Traversa, D., Di Cesare, A., Wright, I., Lacher, D.W., 2016.
250 Updates on feline aelurostrongylosis and research priorities for the next decade. *Parasit.*
251 *Vectors.* 9(1), 389.

252 Elsheikha, H.M., Schunack, B., Schaper, R., 2017. Prevalence of feline lungworm
253 *Aelurostrongylus abstrusus* in England. Abstract Book , 407, The 26th International
254 Conference of the World Association for the Advancement of Veterinary Parasitology,
255 WAAVP, Kuala Lumpur, Malaysia.

256 Garcia-Campos, A., Power, C., O'Shaughnessy, J., Browne, C., Lawlor, A., McCarthy, G.,
257 O'Neill, E.J., de Waal, T., 2018. One-year parasitological screening of stray dogs and cats
258 in County Dublin, Ireland. *Parasitology.* Dec 18:1-7. doi: 10.1017/S0031182018002020.
259 [Epub ahead of print]

260 Genchi, M., Ferrari, N., Fonti, P., De Francesco, I., Piazza, C., Viglietti, A., 2014. Relation
261 between *Aelurostrongylus abstrusus* larvae excretion, respiratory and radiographic signs
262 in naturally infected cats. *Vet. Parasitol.* 206 (3/4), 182-187.

263 Gerichter, C.B., 1949. Studies on the nematodes parasitic in the lungs of Felidae in Palestine.
264 *Parasitology* 39, 251-262.

265 Giannelli, A., Capelli, G., Joachim, A., Hinney, B., Losson, B., Kirkova, Z., René-Martellet,
266 M., Papadopoulos, E., Farkas, R., Napoli, E., Brianti, E., Tamponi, C., Varcasia, A.,
267 Margarida, Alho A., Madeira de Carvalho, L., Cardoso, L., Maia, C., Mircean, V.,
268 Mihalca, A.D., Miró, G., Schnyder, M., Cantacessi, C., Colella, V., Cavalera, M.A.,
269 Latrofa, M.S., Annoscia, G., Knaus, M., Halos, L., Beugnet, F., Otranto, D., 2017.
270 Lungworms and gastrointestinal parasites of domestic cats: a European perspective. *Int.*
271 *J. Parasitol.* 47, 517-528.

272 Giannelli, A., Ramos, R.A., Annoscia, G., Di Cesare, A., Colella, V., Brianti, E., Dantas-
273 Torres, F., Mutafchiev, Y., Otranto, D., 2014. Development of the feline lungworms

274 *Aelurostrongylus abstrusus* and *Troglostrongylus brevior* in *Helix aspersa* snails.
275 Parasitology 141, 563-569.

276 Grabarević Ž., Ćurić S., Tustonja A., Artuković B., Šimec Z., Ramadan K., Živičnjak T.
277 1999. Incidence and regional distribution of the lungworm *Aelurostrongylus abstrusus* in
278 cats in Croatia. Veterinarski Arhiv 69, 279-287.

279 Gunn-Moore, D., Elsheikha, H.M., 2018. Current status of feline lungworm in the UK. Vet.
280 Rec. 182(4), 113-114.

281 Hamilton, J.M., 1968. Studies on re-infestation of the cat with *Aelurostrongylus abstrusus*. J.
282 Comp. Pathol. 78(1), 69-72.

283 Hansen, A.P., Skarbye, L.K., Vinther, L.M., Willesen, J.L., Pipper, C.B., Olsen, C.S., Mejer,
284 H., 2017. Occurrence and clinical significance of *Aelurostrongylus abstrusus* and other
285 endoparasites in Danish cats. Vet. Parasitol. 234, 31-39.

286 Knaus, M., Kusi, I. Rapti, D., Xhaxhiu, D., Winter, R., Visser, M., Rehbein, S., 2011.
287 Endoparasites of cats from the Tirana area and the first report on *Aelurostrongylus*
288 *abstrusus* (Railliet, 1898) in Albania. Wien. Klin. Wochenschr. 123, 31-35.

289 Mundhenke, H., Dauschies, A., 1999. Studies on the prevalence of endoparasites in cats in
290 Hannover and surroundings. Wien. Tierarztl. Monatsschr. 86 (2), 43-48.

291 Olsen, C.S., Willesen, J.L., Pipper, C.B., Mejer, H., 2015. Occurrence of *Aelurostrongylus*
292 *abstrusus* (Railliet, 1898) in Danish cats: A modified lung digestion method for isolating
293 adult worms. Vet. Parasitol. 210, 32-39.

294 Penagos-Tabares, F., Lange, M.K., Chaparro-Gutiérrez, J.J., Taubert, A., Hermosilla, C., 2018.
295 *Angiostrongylus vasorum* and *Aelurostrongylus abstrusus*: Neglected and underestimated
296 parasites in South America. Parasit. Vectors 11, 208.

297 Pennisi, M.G., Hartmann, K., Addie, D.D., Boucraut-Baralon, C., Egberink, H., Frymus, T.,
298 Gruffydd-Jones, T., Horzinek, M.C., Hosie, M.J., Lloret, A., Lutz, H., Marsilio, F.,

299 Radford, A.D., Thiry, E., Truyen, U., Möstl, K.; European Advisory Board on Cat
300 Diseases., 2015. Lungworm disease in cats: ABCD guidelines on prevention and
301 management. J. Feline Med. Surg. 17, 626-636.

302 Robben, S.R., le Nobel, W.E., Döpfer, D., Hendriks, W.M., Boersema, J.H., Fransen, F.,
303 Eysker, M.E., Infections with helminths and/or protozoa in cats in animal shelters in the
304 Netherlands.2004. Tijdschr Diergeneeskd. 129(1), 2-6.

305 Scott, D.W., 1973. Current knowledge of aelurostrongylosis in the cat. Literature review and
306 case reports. Cornell Vet. 63, 483-500.

307 Traversa, D., Lia, R.P., Boari, A., Di Cesare, A., Capelli, G., Milillo, P., Otranto, D.,
308 2009. Feline aelurostrongylosis: epidemiological survey in central and southern Italy.
309 Veterinaria 23, 41-45.

310 Traversa, D., Lia, R.P., Iorio, R., Boari, A., Paradies, P., Capelli, G., Avolio, S., Otranto, D.,
311 2008. Diagnosis and risk factors of *Aelurostrongylus abstrusus* (Nematoda, Strongylida)
312 infection in cats from Italy. Vet. Parasitol. 153, 182-186.

313 Zottler, E.M., Bieri, M., Basso, W., Schnyder, M., 2018. Intestinal parasites and lungworms
314 in stray, shelter and privately owned cats of Switzerland. Parasitol. Int. 69, 75-81.

315 Zottler, E.M., Strube, C., Schnyder, M., 2017. Detection of specific antibodies in cats
316 infected with the lung nematode *Aelurostrongylus abstrusus*. Vet Parasitol. 235, 75-82.

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324 **Table 1.**

325 The prevalence of *A. abstrusus* across seven administrative regions in England.

Geographic region	Prevalence*	Odd ratios	95% CI	p-value
Greater London	1/17 (5.8)	9.632	0.43 to 84.05	0.07
South East	4/72 (5.5)	7.68	1.70 to 32.76	0.01
West Midlands	3/69 (4.3)	6.2	1.21 to 26.84	0.02
North West	2/57 (3.5)	4.25	0.59 to 20.89	0.09
South West	1/46 (2.1)	2.48	0.12 to 17.64	0.43
North East	1/49 (2.0)	1.88	0.10 to 12.24	0.57

326 * Using East Midlands as a reference [5 infected out of 623 tested (80.2%)], Greater London,
327 North West, South East, West Midlands showed significant differences. Prevalence is
328 shown as number of positive samples/total number tested (%).

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342 **Table 2.**

343 Lifestyle distribution and positivity rates of *A. abstrusus* for cats examined in this study.

344 Correlation was established only between cats living indoor and *A. abstrusus* infection.

Lifestyle category	No. of uninfected cats	No. of infected cats	Larvae per gram of feces
Indoor	375	1	12.0
Outdoor Access	455	13	11.5
Stray	81	3	12.0
Feral	22	0	0.0
Total (<i>n</i> = 950)	933	17	11.6

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