New evidence for the age and palaeoecology of the Knysna Formation, South Africa

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Abstract

The existence of lignitic deposits (Knysna Formation) on the South African south coast, near to the town of Knysna has been recognised for more than a century. However, a combination of limited study and few natural exposures has meant that the age and stratigraphic position of the Knysna Formation are unclear, despite its potential as a palaeoenvironmental archive. We present a new suite of chronological, geochemical and palynological data obtained from a recently identified lignite exposure in this area. The lignite pollen assemblage is dominated by palms (Arecaceae), which are now locally extinct, and contains additional palynomorphs of tropical affinity, along with (moisttemperate) Podocarpus-type pollen, grasses, and herbaceous pollen types (e.g. Cliffortia-type, Asteraceae). Overall, the assemblage shows some commonalities with the Miocene Elandsfontein Formation in the Western Cape. The lignites are dominated by a diverse range of higher plant biomarkers, including abundant leaf wax lipids, as well as lignin monomers and leaf cuticle-derived macromolecular organic matter. All strongly indicate a terrestrial depositional setting, perhaps akin to contemporary palm swamps. A number of sesquiterpenoids imply the presence of gymnosperms, supporting observations from the pollen data and previously reported macro-fossil finds. The application of isothermal thermoluminescence techniques to coversands overlying the lignite produced a minimum age of 1.7 Ma. Additional clues as to the likely age of the lignite are provided by compound-specific stable carbon isotope analyses of the leaf wax lipids. These are approximately 8 % enriched relative to typical C₃ vegetation leaf waxes, and imply a potential contribution from C₄ vegetation. From this, an age post-dating the Oligocene may be inferred, and in conjunction with the site's geomorphic setting, an age post dating the middle Miocene is considered plausible. This is

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Keywords: Neogene, pollen, biomarker, luminescence, stable isotope

markedly younger than previous (Eocene) age estimates for the Knynsa Formation.

Due to its progressive aridification and predominantly erosive landscape history

Introduction

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(Partridge and Maud, 1987) southern Africa suffers from a paucity of palaeoecological evidence. The Neogene palaeoecology of the southern African flora is however of great significance and relevant to wider questions concerning patterns of floral dispersal and diversification (Coetzee, 1983; Linder, 2003), the causes and effects of the Late Neogene aridification of southern Africa, and the emergence of C₄ grasslands (Ségelan et al., 2007). The occurrence of organic-rich sediments ("lignites") near to Knysna on the southern Cape coast has been known of for more than a century (Phillips, 1927 and references therein). These are now formally recognised as the Knysna Formation (Jacobs 1992), and comprise sands, fine-grained muddy lignitic sands and lignites up to 30 m thick. Their relative antiquity and palaeoecological significance were easily recognised (Phillips, 1927; Du Toit, 1956). Subsequently, Thiergart et al. (1963) and Coetzee et al. (1983) reported palynological assemblages significantly different from the region's contemporary vegetation. These studies revealed the presence of extant temperate forest vegetation (e.g. Podocarpus sp., Widdringtonia cupressoides), the absence of a number of common contemporary palynomorphs (e.g. Ericaceae), and the presence of locally extinct flora (e.g. Nypa-type palm). The extent, age and stratigraphic relationships between exposures of the Knysna Formation remain unclear, and detailed information such as sample site location has often been lacking in previous reports (e.g. Helgren and Butzer, 1977; Coetzee et al., 1983; Thwaites and Jacobs, 1987). The limited available exposure and borehole data indicate that

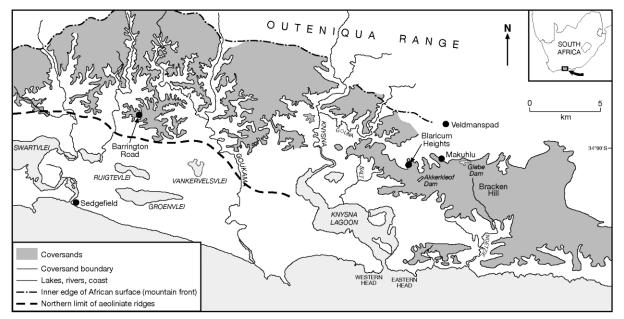
all deposits are concentrated in the Veldmanspad and Brakenhill areas on a platform at

altitudes more than 200 m above modern sea-level (**Figure 1**; Jacobs, 1992). This landscape setting alone implies an antiquity considerably greater than the Quaternary aged dunes and wetlands immediately seawards (Holmes et al., 2007). Age estimates based on palynological evidence (Coetzee et al., 1983) or landscape setting/stratigraphic relationships (Butzer and Helgren 1972; Helgren and Butzer 1977; Thwaites and Jacobs, 1987; Marker and Holmes, 2002; Holmes et al., 2007) are disparate, encompassing a range between the Eocene and the Miocene.

In the course of recent investigations into the origin and distribution of the Knysna coversands, Holmes et al (2007) reported a 2-3 m thick sandy lignite deposit beneath a 15 m sequence of aeolian coversands at Makhulu Quarry, c. 4 km southeast of Veldmanspad (**Figures 1 and 2**). Optically stimulated luminescence (OSL) dating of the coversands produced a minimum age of > 500 ka, but the underlying lignite was not investigated. The aims of the present paper are:

- Following initial attempts at OSL dating (Holmes et al., 2007), to reassess the age of the coversands at Makhulu Quarry using recently developed isothermal thermoluminescence (ITL) techniques. (Choi et al., 2006; Jain et al., 2007; Vandenberghe et al., 2009).
- 2. To analyse the organic geochemistry and palaeoecology of the lignite exposed at Makhulu in order to infer its likely depositional context and palaeoenvironment/palaeoecology.
- 3. To set the palaeoecological data in the context of palynological (Cole and Roberts, 1996; Scott, 1995) and palaeoecological (Hopley et al., 2007; Ségelan et al., 2007) evidence for southern African flora.

Figure 1: Site locations in the Wilderness-Knysna region shown in relation to the mapped coversand distribution (modified from Marker and Holmes, 2002). The broad locations of previous lignite exposures (Brakenhill and Veldmanspad) are also shown.



Site description and stratigraphic setting

Samples were obtained from Makhulu Quarry in 2004 (34°00′48″S 23°06′26″E; altitude 220 m above modern sea-level [amsl]) which lies northeast of Knysna, and close to the northern limits of the Knysna coversands mapped by Marker and Holmes (2002). The stratigraphy of the site (which since sampling has been fully excavated) is shown in **Figure 2**. The site lies within a hollow in the underlying Peninsula Formation (Table Mountain Group) bedrock topography, which probably accounts for the enhanced thickness of the sand in this location (Holmes et al., 2007). Up to 3 m of sandy lignites unconformably overlie the Peninsula Formation quartzites and are themselves overlain by up to 15 m of weathered and ferrugenised coversand deposits. Holmes et al. (2007) considered this the southern Cape coversand type site and provided a detailed description of the coversand properties.



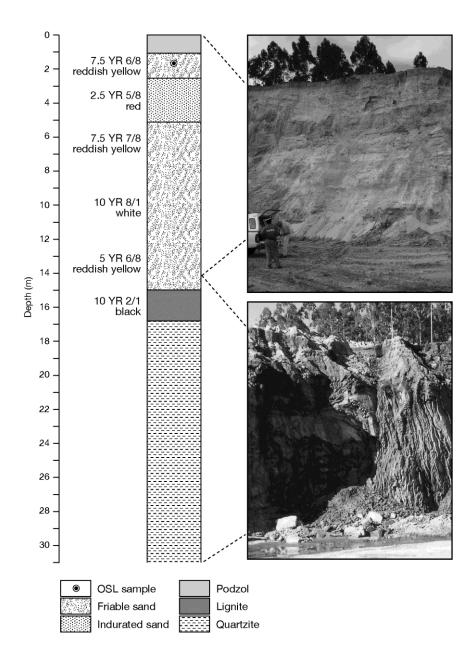


Figure 2: Site stratigraphy and photographs of Makhulu Quarry (in 2004). The location of the OSL sample reported by Holmes et al., (2007) and re-evaluated in this study is shown. Note in upper photograph the laterisation of the coversands (at c. 3-5 m depth) and in the lower photograph the lignite/coversand contact.

Geochronology

Optically stimulated luminescence (OSL) of quartz has been widely used in the southern Cape to date mid to late-Pleistocene marine and aeolian sediment (e.g. Bateman et al., 2004). The fundamental principle is that buried quartz acts as a dosimeter, storing up trapped electrons in its crystal lattice in proportion to the natural (geological) radioactivity in the environment (Aitken 1985). If buried for sufficient time quartz grains become 'saturated' (lattice cannot accommodate additional free electrons), which constrains the upper limit of the technique. The time to saturation is largely a function of background radioactivity and in low radiation environments reliable OSL ages approaching 500 ka have been determined (Banerjee et al., 2003). Holmes et al. (2007) found that the coversands in the Makhulu Quarry and the nearby Blaricum Heights site (Figure 1) were saturated using standard OSL techniques and obtained minimum ages of >500 ka and >300 ka respectively (Figure 2).

In this study we re-analyse one of the Makhulu samples reported by Holmes et al. (2007), using an isothermal thermoluminescence (ITL) dating approach which seeks to overcome the saturation issues previously encountered (Choi et al., 2006). While most OSL dating relies on measuring electrons immediately evicted on optical stimulation (the 'fast component' of the luminescence signal; Wintle and Murray, 2006), quartz grains also emit a thermoluminescence (TL) signal upon heating, with peaks at 325°C and 375°C (Aitken, 1985). New ITL dating protocols utilise a single aliquot regenerative dose (SAR) protocol akin to standard OSL dating, but measure only the signal emitted from the 325°C TL peak, which is known to be bleached by visible light (Franklin, 1997). Crucially, this signal saturates at much higher doses than the OSL fast component. For instance, Jain et al. (2005) recorded a near

order of magnitude increase in the saturation dose of quartz grains using ITL compared to OSL. There is thus potential for ITL to be used to date sediments beyond the range of conventional OSL dating, which would be of significance given the extensive Neogene marine and aeolian deposits along the Western Cape coastline.

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Methods

Luminescence sampling, quartz sample extraction and dose rate determinations followed Holmes et al. (2007). All luminescence measurements were carried out on a Risø luminescence reader using 9.6 mm diameter aliquots with luminescence signals detected through a Hoya U340 filter. Previous studies have employed a range of temperatures for both the ITL stimulation and the sample preheat (Choi et al., 2006; Buylaert et al., 2006; and Huot et al., 2006). To establish the applicability of ITL to the deposits in this study the protocol was first tested using a coversand sample (Shfd08191 from Barrington Road, 33° 59'13 S, 22° 51'08 E; 90 m amsl; Figure 1) which was within the range of the standard SAR OSL approach (equivalent dose of c. 240 Gy). For this sample SAR OSL equivalent doses (D_e) were determined following Murray and Wintle (2000), with ITL De values at both 300°C and 310°C determined for a range of preheat temperatures (240°C to 270°C in 10°C increments; Choi et al., 2006; **Table 1**). For sample Shfd08191 ITL Des obtained at 310°C were consistently closer to the OSL De than those obtained using ITL at 300°C (Figure 3). The measurements at 310°C also produce a D_e plateau at preheats of 260°C and below. Based on these results, an ITL stimulation temperature of 310°C and a preheat temperature of 260°C were selected for full ITL measurement of the sample. Growth curves were fitted with a saturating exponential function (Figure 4). For the purposes of subsequent age estimation

all aliquots with a recycling ratio outside 1 \pm 0.2 were rejected prior to D_e calculation. Sunlight bleaching of the ITL signal was tested by exposing seven aliquots of the Makhulu Quarry sample (Shfd04287) to (UK) daylight for 1 week. A residual ITL D_e of 4.73 \pm 0.34 Gy (n=7) was determined (**Figure 5a**; **Table 2**), which given the magnitude of the sample equivalent doses, was considered small enough to be of negligible significance. For the Makhulu sample (Shfd04287) 24 aliquots were measured using an ITL stimulation temperature of 300°C and a preheat temperature of 260°C (determined experimentally in an identical manner to the above). **Table 2** shows the finalised OSL and ITL D_e values used for age calculation reported with 1 sigma uncertainties.

Results

For Shfd08191 (Barrington Road), the mean D_e values from the OSL and ITL approaches are broadly similar and are within 1 standard deviation of each other (OSL D_e = 242 \pm 30; ITL D_e = 295 \pm 50; **Figure 5**). The slightly higher mean D_e for ITL results may reflect a signal contribution from TL traps incompletely bleached by sunlight prior to burial (**Figures 5b and 5c**). More significantly in the context of the coversands, while the OSL growth curves typically saturate at 350 \pm 50 Gy (**Figure 4a**), the ITL growth curves grow beyond 550 Gy (**Figure 4b**). As such, and given the favourable comparison of ITL to OSL data, it is inferred that the ITL procedure can provide new age constraints on Knysna coversand material beyond those presented by Holmes et al (2007). ITL measurements on the Makhulu quarry sample (Shfd04287) reveal the naturally acquired ITL signal to be very large (**Figure 6**) and for two aliquots this was significantly higher than the maximum administered laboratory dose, which prevented interpolation. These aliquots were rejected from

subsequent analyses. The remaining aliquots were fitted with saturating exponential growth curves (**Figure 6b**), of which eight were in saturation (on average the Makhulu sample growth curves saturated at 1390 ± 490 Gy). The remaining 10 aliquots, for which a D_e could be estimated, produced a mean D_e of 806 ± 58 Gy (**Figure 5d**), from which an age of 1.7 ± 0.14 Ma was derived (**Table 2**). With 40% of the aliquots measured by ITL in saturation this is clearly a minimum D_e and thus, age estimate for the coversand (and lignite) at this site. It is however more than triple the minimum age obtained via SAR OSL (Holmes et al., 2007) and it suggests a pre-Quaternary antiquity for both the coversands and the underlying lignite.

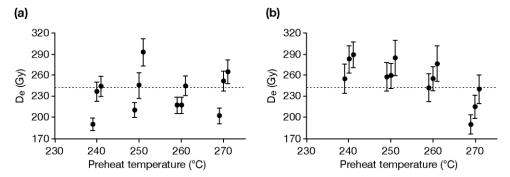


Figure 3: Preheat plateau tests undertaken to establish the optimum ITL stimulation and preheat temperatures for sample Shfd08191: (a) ITL at 300° C (b) ITL at 310° C. Dashed line shows D_e of sample established using conventional SAR OSL. Note for clarity aliquots measured at the same preheat temperatures are slightly offset.

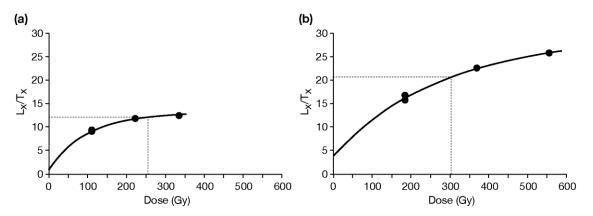


Figure 4: Example growth curves for individual aliquots from sample Shfd08191: (a) OSL and (b) ITL

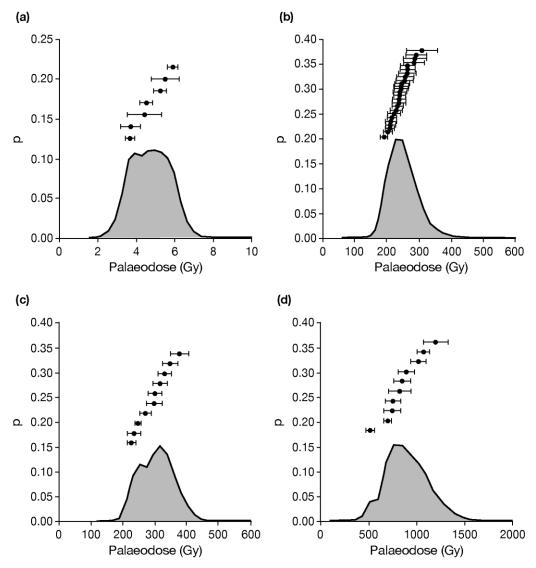


Figure 5: D_e distributions of samples measured (a) ITL of sample Shfd04287 after exposure to daylight for seven days. Average D_e = 4.73 \pm 0.34 Gy (b) OSL D_e distribution for sample Shfd08191 from Barrington Road (c) ITL D_e distribution for sample Shfd08191 from Barrington Road (d) ITL D_e distribution for sample Shfd04287 from Makhulu Quarry.

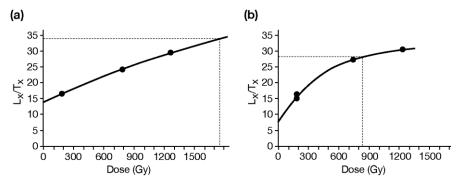


Figure 6: Example ITL growth curves for sample Shfd04287

Organic geochemistry

The last two decades have seen the rapid development of palaeoecological and palaeoenvironmental approaches using biomarkers (see Eglinton and Eglinton, 2008 for an overview). Suites of biomarkers within organic matter reflect the composition of the original inputs to a sample (e.g. local ecology) and the effects of subsequent microbial and diagenetic degradation. In this study we have applied solvent-extractable lipid analyses, and pyrolysis-GC/MS (py-GC/MS) analyses to the Makhulu lignite. The pyrolysis technique provides insights into the origin and diagenetic alteration of insoluble macromolecular organic matter (e.g. lignin) within a sample. Using these techniques we assess the Makhulu depositional setting, the extent of organic matter preservation/diagenesis, and the palaeoenvironmental setting of the site. These new data are interpreted in light of accompanying palynological analyses, and potential points of comparison for future lignite finds are noted.

Methods

The sampled lignite was freeze dried and homogenised prior to analysis. Total C and N data were measured using a SerCon ANCA GSL elemental analyser connected to a SerCon

Hydra 20-20 continuous flow isotope ratio mass spectrometer (cf-IR-MS). Solventextractable lipid biomarkers were extracted using a series of solvents of progressively decreasing polarity (methanol; dichloromethane (DCM); hexane). Extracts were eluted over an Al₂O₃ column to derive apolar and polar fractions. The polar fractions were derivatized using bis(trimethylsilyl)trifluoroacetamide (BSTFA) for 1 hour at 60°C prior to analysis. GC/MS analyses were carried out on a Perkin Elmer Clarus 500 GC-MS system fitted with a CP-Sil 5CB MS column (30 m x 0.25 mm). Compounds were identified on the basis of their retention times and mass spectra. Compound specific stable isotope analyses were conducted on the apolar solvent fraction using an Agilent 6890N gas chromatograph equipped with a combustion reactor, interfaced to a SerCon GC-CP which was connected to the Hydra 20-20 cf-IR-MS. Results are presented as an average of three solvent injections with accompanying standard deviations (better than 0.5 % for all reported compounds). Pyrolysis GC/MS was conducted using a CDS 1000 pyroprobe coupled to the Perkin Elmer Clarus 500 GC-MS system. Approximately 0.5 mg of sample was encapsulated in a quartz tube and held in place with quartz wool. The sample was then pyrolysed at 610°C for 15 seconds. Measurements were conducted on an untreated sample, as well as the residue remaining following saponification (reflux in KOH and methanol). This process hydrolyses certain chemical bonds (ester bonds), and assists in characterising the original nature of the macromolecular organic matter (some macromolecules will be resistant to this process).

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Results

The total organic carbon (TOC), total nitrogen (TN) and $\delta^{13}C_{TOC}$ of the lignite are shown in **Table 3**. The TOC/TN ratio is very high (45) and suggestive of organic matter

dominated by terrestrial / higher plant-derived organic matter, as opposed to algal-rich lacustrine organic matter, which typically exhibits lower TOC/TN ratios (4-10; Meyers and Lallier-Vergès, 1999). These data are consistent with Phillip's (1927) observations of well-preserved plant macro-fossils within the lignite.

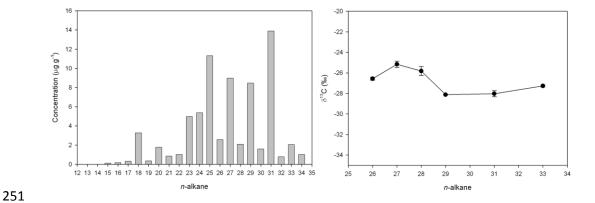
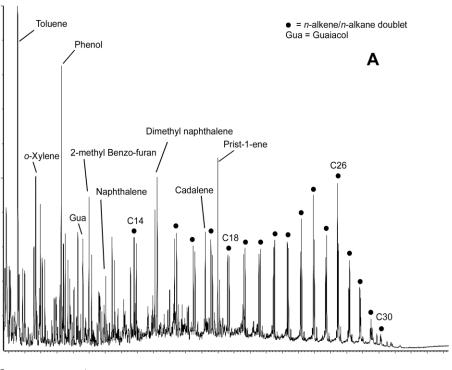


Figure 7: The *n*-alkane distribution (absolute concentrations) and *n*-alkane δ^{13} C values obtained from the apolar solvent extract. End member *n*-alkane δ^{13} C values for C₃ and C₄ vegetation are typically - 20‰ and -35‰ respectively (Boom et al., 2002).



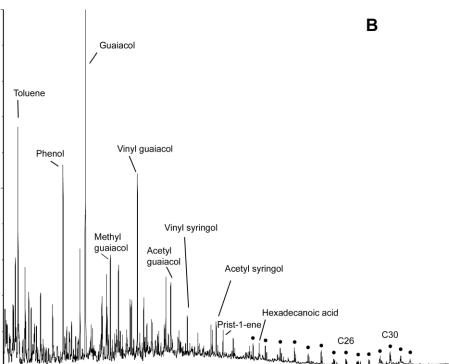


Figure 8: Pyrograms (total ion current) of (A) Knysna lignite and (B) contemporary wetland sediments from Still Bay area. Peak heights are normalised to the highest peak in each sample. The GC programme comprised an initial temperature of 40°C for 1.8 minutes, ramped to a final temperature of 310°C at 4°C min⁻¹ which was held for 20 minutes. Major peaks are labelled (with chain length indicated where appropriate) with selected structures shown

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The extractable lipids comprise a homologous series of *n*-alkanes, pentacyclic triterpenoids and des-A-triterpenoids (des-A-Lupane and Olean-12-ene) in the apolar extract, and a homologous series of n-alkanols (C_{16} - C_{28}), as well as the compounds β -amyrin (olean-12-en-3 β -ol), α -amyrin and friedelan-3-one in the polar extracts. The *n*-alkane distribution (Figure 7) is bi-modal, peaking at C25 and C31. n-alkanes in the range C27 to C33 are generally considered to derive from higher plant epicuticular leaf waxes (Eglinton and Hamilton, 1967) and as observed here, are characterised by odd over even chain length dominance (expressed as the carbon preference index; 'CPI'). A secondary peak is seen within the mid chain length n-alkanes (C_{21} to C_{25}). These shorter odd chain length n-alkanes (C₂₃ and C₂₅) have been associated with submerged or floating plants, rather than terrestrial plants or emergent macrophytes such as Typha and Phragmites (Ficken et al., 2000). This distinction forms the basis of the P_{aq} index, which is used to assess the relative significance of submerged, emergent or terrestrial vegetation (Ficken et al., 2000). The P_{aq} value of 0.42 for the Makhulu lignite is transitional between terrestrial vegetation / emergent macrophytes (typically 0.1-0.4) and submerged / floating plant matter (typically 0.4 - 1.0). This interpretation implies the presence of some floating/aquatic vegetation, and potentially a partially open water environment.

A number of sesquiterpenoids, notably caladene, and calamenene were detected (Bechtel et al., 2002; Stefanova et al., 2005). These are typical coal biomarkers and are common constituents of resins from the Podocarpaceae, Cupressaceae and Pinaceae families (Otto et al., 2005). In this case the sesquiterpendoids extracted from Makhulu cannot be associated with any individual plant family, but their occurrence strongly suggests the presence of gymnosperm vegetation in the site vicinity. Similarly, although most lipid

biomarkers are indicative of a higher plant (angiosperm) origin, none can be used as chemotaxonomic indicators (e.g. Pancost and Boot, 2004).

The other major constituents of the lipid extracts are hopanes, and their diagenetic isomers, moretanes, which are typically derived from bacterial activity and are common in many lignite deposits (e.g. Bechtel et al., 2002; Bechtel et al., 2003; Otto et al., 2005). Here they are dominated by 17β -22,29,30 trisnorhopane and 17β 21 α -homomoretane. Hop-17(21)-ene is also present. Diagenetically immature samples are dominated by hopanes in the (biological) 17β 21 β isomeric configuration (Pancost et al., 2007) and the presence of some hopanes in the 17α 21 β configuration reveals the occurrence of minor diagenetic alteration.

The lignite pyrolysate (i.e. solvent insoluble OM) is shown in **Figure 8A**, with surface sediment from a contemporary southern Cape wetland for comparison (**Figure 8B**). The Makhulu lignite is dominated by aromatic and aliphatic compounds, with the latter, which comprise a series of long chain (C_8 to C_{32}) n-alkene/n-alkane doublets, accounting for 52% of the total ion current (TIC) and peaking at chain lengths of C_{10} - C_{15} , and C_{21} to C_{26} . These aliphatics are substantially more significant in the lignite than in the modern sample (< 19% of TIC). They may be derived from a range of potential biological sources, including cutin, a biopolymer derived from leaf cuticles, which may be preserved in samples of a geological antiquity (Logan et al. 1993). Other potential sources include cutan, a resistant biopolymer derived from certain plant cuticles (Nip et al., 1989; Boom et al., 2005), algaenan, which is derived from some freshwater algae (Blokker et al., 1998), and geo-polymers formed post-deposition (Yang et al., 2005; Gupta et al. 2007). The pyrolysate of the saponified residue reveals a reduction in the significance of the doublets longer than C_{24} , implying the presence

of cutin monomers (Logan et al., 1993). A residual aliphatic signal remains, and given the lack of evidence for algal organic matter in the sample (e.g. high TOC/TN ratio, the low significance of short chain alkanes, and the absence of specific algal biomarkers) this most likely represents a plant-derived macromolecule (e.g. cutan) and/or a post-depositional geopolymer. The former source is of interest given the isolation of cutan in the leaves of extant *Podocarpus* sp. (Boom et al., 2005), and the occurrence of macro-remains of *Podocarpus* in the Knysna Formation (Phillips, 1927). In conjunction, a number of lignin pyrolysis products were identified, which are of a lower relative significance and diversity (0.8% TIC) than the modern sample (c. 30% of TIC). This probably reflects the aerobic degradation of lignin. The lignin monomers are dominated by guaiacol and degradation is implied by both the lack of alkylated guaiacol lignin monomers and the trace amounts of (less resistant) sinapyl-derived lignin monomers.

Compound specific stable isotopes

Inferring terrestrial palaeoenvironmenta from bulk stable isotope data in lacustrine contexts may be complicated by the incorporation of biomass from multiple sources (Street-Perrott et al., 2004). Due to their wide distribution within the atmosphere (Conte and Weber, 2002) the isotopic signature of leaf wax lipids can provide useful insights into regional palaeoecological conditions (Boom et al., 2002). The δ^{13} C values determined for the C₂₇ to C₃₃ n-alkanes in the Makhulu lignite fall between -25 and -28 ‰ (weighted mean δ^{13} C -27.0 ‰; **Figure 7 and Table 4**). The dominance of the C₃₁ alkane and the depletion of the C₂₉ and C₃₁ homologues are consistent with a leaf wax origin (Collister et al., 1994). These

isotopic ratios are substantially more enriched than leaf waxes derived from pure C_3 vegetation (typical $\delta^{13}C$ of -35 %; Collister et al., 1994; Boom et al., 2002).

Compound specific δ^{13} C measurements on the bacterially-derived hopanes (**Table 4**) reveal slight enrichment (-20 and -25 ‰) relative to the bulk organic matter (-26.6 ‰). This has been observed in both contemporary wetlands and Palaeogene lignites (Pancost et al., 2007). The hopane δ^{13} C values do not indicate anaerobic conditions (*ibid*), which is consistent with the py-GC/MS data for the (aerobic) degradation of lignin. Similarly, there is no evidence for anaerobic methane oxidation (e.g. depleted hopane δ^{13} C values).

Palynological analysis

Palynomorphs (Figure 9 and Table 5) are abundant in the Makhulu lignite, but are often poorly preserved (8% were damaged or unidentifiable). From a count of 250 palynomorphs the sampled assemblage is dominated by palms (37 %). Poaceae (4.6 %,) Cyperaceae (6.7 %) and *Podocarpus*-type pollen (3.8 %) are also frequently identified. The palm (Arecaceae) pollen is mostly monosulcate in form, preventing easy associated with a single family or subfamily (e.g. Harley and Baker, 2001). Pollen from typical *fynbos* vegetation is mostly lacking (e.g. Ericaceae). One palynomorph closely resembling the Rosaceae genus *Cliffortia* was identified, along with pollen resembling *Anthospermum*-type (Rubiaceae) and two examples of a small tricolporate palynomorph akin to Grubbiaceae. One triporate palynomorph with a triangular amb potentially represents Proteaceae. Asteraceae pollen (2.9 %) of the Vernonia and Asteroideae types was identified (e.g. Scott,

1982; Skvarla et al., 2005). A number of palynomorphs were of unclear affinity, the most common being a tetracolporate grain (6.7 %), resembling pollen from the family Sapotaceae (Figure 9), and a tetraporate grain (5 %) resembling the pollen of *Symphonia* (Clusiaceae; Figure 9). Pollen resembling the latter type has also been identified in Miocene sediments from the Angola Basin (Partridge, 1978). In the contemporary tropics (South America, West Africa, and Madagascar) this genus is often riparian or found within palm swamps (Ledru, 2001; Berrio et al., 2002). Along with palms, other pollen types of potential tropical affinity include Malvaceae and Moraceae. Trilete and monolete spores were also common, and comprised 3.8 % of the pollen spectrum.

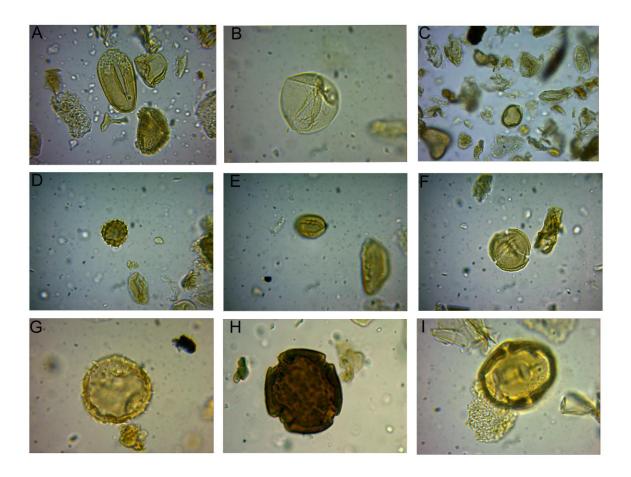


Figure 9: Photograph montage of key pollen types identified within the sample A: Palm; B: Poaceae; C: *Cliffortia*-type; D: Asteraceae; E: *Rhus*-type; F: Rubiaceae(?); G: Malvaceae; H: *Symphonia*-type; I Tetracolporites (Sapotaceae).

Discussion and synthesis

Depositional setting

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Overall, the data imply that the lignite formed in a marshy environment surrounded by significant emergent and riparian vegetation (e.g. Cyperaceae pollen). The biomarker data and the bulk C/N ratios indicate little algal organic matter contribution to the lignite. The pyrolysis data similarly imply that a substantial proportion of the macromolecular organic matter is derived from terrestrial plants; notably leaf cuticles (cutin) and lignin monomers. The presence of the sesquiterpenoids cadalene and calamenene in the lipid extracts is strongly indicative of inputs from gynomsperm resins and is supported by the presence of *Podocarpus* pollen. The biomarker data are therefore primarily indicative of inputs from terrestrial/riparian plant-derived organic matter, although the Pag parameter implies the presence of some submerged or aquatic plants. Such a terrestrial wetland setting is consistent with some interpretations of the Knysna Formation (Coetzee et al., 1983). This was not a strongly anoxic environment, nor is there evidence for marine-derived organic matter (c.f. Volkman et al., 2008). This does not rule out wetland formation associated with high water tables during periods of high relative sea-level (sensu Cole and Roberts 1996), but it is not strongly supportive of a back-barrier lagoon or estuarine setting. The difficult terrain and limited exposure means that the stratigraphic relationship between Makhulu and other reported lignite exposures in the Knysna region (i.e. Veldmanspad and Brakenhill) is uncertain (Holmes et al., 2007). Such complexity serves to highlight the need to document the biostratigraphic and palaeoecological evidence within each lignite exposure. The maximum recorded altitude of the lignites is 320 m amsl (Veldmanspad), with the Brakenhill lignites reportedly lying at c. 259 m amsl (Thwaites and

Jacobs, 1987) and Makhulu at 220 m amsl. Marker and Holmes (2002) noted the altitudinal and spatial separation of the Knysna Formation deposits at Veldmanspad, Brakenhill and Fisantshoek from the coversands, while Thwaites and Jacobs (1987) also suggested that lignite deposits may represent spatially and temporally discrete deposits. This clearly has implications for age-related and palaeoecological comparisons with the Makhulu lignite, but it is hoped that the palynological and geochemical data presented here will provide useful points of comparison.

Age estimates

The ITL analyses on the overlying coversand provide the first numerical age constraints on the Knysna Formation. While the data presented by Holmes et al (2007) demonstrated an age pre-dating the Late Pleistocene, the application of ITL pushes this minimum age back to > 1.7 Ma. The gradational boundary between the coversand and lignite units suggests that they were deposited at a similar time. The ITL result therefore implies that the Makhulu lignite may pre-date the Quaternary Period. The ITL age of 362 ± 35 ka for the Barrington Road coversand (Shfd08191) is similar to the minimum ages reported by Holmes et al. (2007) for the Blaricum Heights coversands. Both sites are clearly within the coversand sheet, but their relationship with Makhulu is unclear. Their essentially mid-Pleistocene ages, in conjunction with the new Makhulu ITL data, do however support the previous inference that parts of the coversand system have been reworked during the Quaternary (Holmes et al., 2007).

The compound-specific stable isotope data also provide potential constraints on the likely age of the Makhulu lignite. The weighted mean (C_{27} - C_{31}) n-alkane δ^{13} C value of -27 ‰

indicates inputs from non- C_3 vegetation sources; the end member $\delta^{13}C$ values for C_3 and C_4 plant leaf waxes being -35 and -20 % respectively (Boom et al., 2002). Presently, in southern Africa C_4 photosynthesis is abundant in both grasses (Poaceae) and sedges (Vogel et al., 1978; Stock et al., 2004). Compound-specific stable isotope data of this type have previously been used to track the emergence of C_4 grasses during the Neogene (Freeman and Calarusso, 2001). A simple two part mixing model with the above-noted end members implies a C_4 contribution to the leaf wax signal as high as 53 %. This approach however assumes no input from CAM (crassulacean acid metabolism) vegetation, which exhibit highly variable leaf wax $\delta^{13}C$ values (ranging between -21 to -29 %; Collister et al., 1994; Bi et al, 2005), and it also assumes that there is little inter-species variability in the ^{13}C offset between whole plant tissue and leaf wax n-alkanes (Freeman and Calarusso, 2001).

There is currently little published data on CAM plant leaf wax δ^{13} C variability, making this issue difficult to resolve. This result is however intriguing. Presently CAM plants are found in the southern Cape area, while grass pollen, which can dominate Quaternary and contemporary pollen spectra, is a relatively minor component of the Makhulu pollen spectrum. The efficacious dispersal of leaf waxes in the atmosphere implies that the n-alkane δ^{13} C integrates a vegetation signal from a substantial land area (Conte and Weber, 2002; Rommerskirchen et al., 2003), and in the absence of other evidence at present, a C₄ contribution of some size represents a parsimonious interpretation. Assuming a nonnegligible contribution of C₄ vegetation, an age close to or post-dating the expansion of C₄ grasses in the Late Miocene (c. 10 Ma; Cerling et al., 1993; Freeman and Calarusso, 2001) might be inferred, with an upper age further constrained by genetic evidence for the origins of C₄ vegetation. Given that this origin probably lies in the Oligocene (Tipple and Pagani,

2007), and for C_4 Cyperaceae, in the Miocene (Besnard et al., 2009), even a small C_4 contribution casts doubt on an Eocene age estimate (cf. Coetzee et al., 1983; Thwaites and Jacobs, 1987). In southern Africa Ségalen et al. (2007) concluded that the C_4 expansion occurred as late as 4-5 Ma; an estimate supported by $\delta^{13}C$ data from speleothems in the Makapansgat Valley in the summer rainfall zone (Hopley et al., 2007).

The Makhulu site lies on a coastal platform thought to have formed after a phase of uplift in the early-mid Miocene (Partridge and Maud, 1987). The early to middle Miocene was a phase of global warmth and associated high eustatic sea-levels (Haq et al., 1987; John et al., 2004; Tripati et al. 2009), representing a plausible origin for this surface. This setting is not therefore inconsistent with a Miocene or younger age, as inferred from the n-alkane δ^{13} C data. Given the current site altitude (220 m), a proposed phase of later Neogene uplift (c. 150-200 m in the Pliocene; Partridge and Maud 1987) places the site close to sea-level, the water table and a potential sediment source for the overlying coversands between the late Miocene and early Pliocene (c. 10-5 Ma). Such an interpretation however ignores the significant eustatic sea-level changes during this period (Haq et al., 1987; Le Roux, 1990; John et al., 2004; Miller et al., 2005) and more recently Roberts and Brink (2002) have argued for a stronger eustatic imprint on the Western Cape coastal record. However, disentangling these eustatic and tectonic factors without additional data is clearly difficult and beyond the scope of this paper.

Regional palaeoecology

The pollen preserved at Makhulu provides some indication of the local and regional ecology. Palms are currently extinct in the southern and Western Cape, and along with the

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presence of palynomorphs of possible tropical affinity (Malvaceae, Moraceae, Sapotaceae, and Symphonia-type) the Makhulu data are suggestive of warmer and more humid conditions than the present. Interestingly, given the efficacious dispersal of saccate pollen, the proportion of *Podocarpus*-type does not suggest a dominant presence in local/regional environment (see also Coetzee et al., 1983), which contrasts with Holocene pollen spectra in this region (Martin, 1968). Rhus-type pollen is moderately-well dispersed (Scott, 1982), and may have been present in the local environment. Asteraceae may represent a wide range of southern African environments, but they are especially dominant in contemporary Karoo and fynbos environments. We thus infer a wetland (Cyperaceae, Aponogetonaceae), associated with a surrounding palm swamp or "gallery forest" vegetation community (palms, Symphonia-type), which in turn was surrounded by a mixed vegetation community comprising arboreal taxa (e.g. *Podocarpus* sp.), herbs (*Cliffortia*-type, *Anthospermum*-type) and grasses. The proportion of grasses is relatively low compared to the contemporary environment (Martin 1968), although these may be under-represented given the potentially high proportion of the locally-derived pollen from the palm swap vegetation.

This assemblage differs from previous descriptions of palynomorph assemblages from the Knysna Formation (Thiergart et al., 1963; Coetzee et al., 1983). A comparative summary is shown in **table 5**. Key differences include the dominance of palm pollen at Makhulu, which was either absent or tentatively identified (respectively) by Thiergart et al. (1963) and Coetzee et al. (1983). An additional sample, apparently from the Veldmanspad area, has been reported to contain high concentrations of palm pollen resembling *Jubaeopsis* and *Nypa* (Helgren and Butzer, 1977; Scott, 1995). Pollen of the distinct *Nypa* morphology was not identified at Makhulu. Van Zinderen Bakker (1984) reported palm

pollen in marine sediments as young as the Late Pliocene north of Namibia, while palm pollen of the same monsulcate morphology as Makhulu Quarry has also been identified in the Miocene Elandsfontein formation (Coetzee and Rogers, 1982). Asteraceae pollen, two types of which were found in the Makhulu assemblage, has not been identified in previous studies of the Knysna Formation. Other studies of the Knysna Formation have also reported Proteaceae pollen (possibly identified at Makhulu) to be common, and grass pollen to be present, but rare (Helgren and Butzer 1977; as at Makhulu). Given these apparent differences in floral composition, the possibility that the Knysna Formation sites are diachronous cannot be ruled out. It is currently difficult to evaluate this further given the limited information on previous site locations and pollen assemblages.

Looking further afield, Coetzee (1983) interpreted proposed Miocene pollen spectra from Noordhoek (Cape Town) and the Elandsfontein Formation near Langebaanweg (Coetzee and Rogers, 1982) as indicative of mixed coniferous forests with tropical elements and varying proportions of palms. The minimum age of the pollen assemblage at Langebaanweg is constrained by overlying Varswater Formation (Franz-Odendaal et al., 2002 and references therein) and contains substantial amounts of palm pollen, as well as *Podocarpus, Widdringtonia*, Casuarinaceae, and Anacardiaceae. From the non-arboreal taxa, Asteraceae, *Cliffortia*-type, Proteaceae, and (possibly) Thymelaeceae and *Stoebe*-type were identified. At Noordhoek Asteraceae pollen was sparse or absent in the (presumed) earlymid Miocene parts of the record (Coetzee, 1983). During the early Pliocene increased abundances of Asteraceae pollen were associated with the loss of tropical pollen types; notably the palms (*ibid*; Coetzee 1986).

Additional data pertaining to the Elandsfontein Formation were presented by Cole and Roberts (1996). Here, samples from borehole LNV4 close to the Berg River contained palynomorphs characteristic of *fynbos* vegetation (e.g. Proteaceae, *Passerina*-type, *Anthospermum*-type), as well as Asteraceae (*Brachylaena*-type and undifferentiated) and Poaceae pollen. The similarities were considered sufficient to warrant association with the Langebaanweg record, although palm pollen was absent from the LNV4 samples. Scott (1995) presented data for three samples from the Pliocene Varswater Formation and identified a range of typical *fynbos* pollen types (*Cliffortia*-type, Restionaceae, Proteaceae, and Ericaceae), along with Asteraceae and grass pollen. Palm pollen was only identified in the deepest sample, which was considered transitional from the sub-tropical environments of the Elandsfontein Formation.

Palaeogene (Eocene) age estimates for the Knysna Formation have been inferred on the basis of the palynological data and notably, the absence of Ericaceae pollen (Thwaites and Jacobs, 1987). It is notable that none of the Knysna Formation sites apparently contain Ericaceae pollen, and nor was it reported at Langebaanweg (Coetzee and Rogers, 1982; Cole and Roberts, 1996). It has however been identified in the Noordhoek borehole where it is becomes progressively more abundant, along with the appearance of Asteraceae pollen in the (assumed) Late Miocene to Pliocene (Coetzee 1983). Ericaceae is also found in the Pliocene Varswater Formation assemblages (Scott, 1995). Asteraceae pollen is found at Makhulu, suggesting potential correspondence with the Noordhoek and the Langebaanweg sites. The Makhulu lignite therefore displays some commonalities with the Langebaanweg pollen assemblages (palm, *Podocarpus*-type, Poaceae, Cyperaceae, Asteraceae, *Cliffortia*-type, Anacardiaceae [*Rhus*-type]), and the lower parts of the Noordhoek record from which

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a similar age, broadly equivalent to the middle Miocene, may be inferred. There are, however, differences and the significant geographic separation of these regions should be borne in mind. Such an age association is not inconsistent with the geomorphic evidence or isotopic evidence at Makhulu. If the leaf wax stable isotope data are indeed influenced by a significant C₄ vegetation contribution, an age more akin to the Late Miocene or younger might be inferred for Makhulu.

A Miocene to Pliocene age range inferred for the Makhulu lignite places its deposition within a key phase of the evolution and diversification of the Cape Flora, a period that apparently records a major shift in the ecology of the Western Cape; viz, the demise of the palms and tropical-affinity vegetation (Coetzee, 1983) and the diversification of many succulent Karoo and fynbos plant lineages (Richardson et al., 2001). A major change around the Late Miocene to Pliocene has been considered indicative of the progressive aridification the Western Cape, which probably related to the ongoing expansion of Antarctic ice (Coetzee, 1983; 1986; Zachos et al., 2001). This situation was also intimately related to the intensification of the Benguela upwelling system (Marlow et al., 2000), which would have reinforced summer atmospheric stability. It is difficult to assess when and how this would have affected the southern Cape/Knysna region, which even today experiences distinctly different climatic conditions to the Cape Town area. A potential C₄ signal in the *n*-alkane stable isotopes implies, to some extent, conditions akin to the contemporary summer rainfall regions of southern Africa (Vogel et al., 1978), and a response to late Neogene reductions in atmospheric pCO₂ (Hopley et al., 2007). Given the pollen data presented here, vegetation of a tropical affinity may have been restricted to the vicinity of wetlands (a 'palm swamp' environment). In conjunction with evidence for an inferred absence of C₄ vegetation on the west coast (Langebaanweg) area during the early Pliocene (Franz-Odendaal et al., 2002) it is probable that distinct vegetation communities in the west coast and southern Cape regions (and most likely contrasting climates) were established during this Late Neogene climatic/ecological re-organisation.

Conclusions

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The lignitic sediments exposed at Makhulu Quarry provide a new window on Neogene palaeoenvironments of the southern Cape coastline. Analyses using the ITL technique provide a revised minimum age of > 1.7 Ma for the Makhulu coversands, suggesting a pre-Quaternary emplacement of the sands and lignite. Additional age constraints on the lignite are provided by the compound specific δ^{13} C data, which imply a potential presence of C₄ vegetation in the regional environment. The organic geochemistry of the lignite is dominated by biomarkers derived from terrestrial plants, with no significant evidence for a marine organic matter contribution. The pollen assemblage is dominated by palms, as well as a riparian vegetation suite that probably surrounded the immediate wetland site. A 'palm swamp' setting surrounded by vegetation comprising a mix of shrubs and grasses and some arboreal species (e.g. Podocarpus sp.) is inferred. These data show marked contrasts with some previous descriptions of palynology of the Knysna Formation, but show some commonalities with pollen assemblages found in the Miocene Elandsfontein Formation in the Western Cape. From this, a probable age range between the Miocene and 1.7 Ma is therefore proposed for the Makhulu lignite, which might be further refined to a post late Miocene age should it prove possible to demonstrate that the n-alkane δ^{13} C signal is dominated by a C₄ leaf wax contribution.

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Table Captions

Table 1: Details of the SAR ITL protocol used in this study (following Choi et al. 2006). All heating was carried out at 2° C s⁻¹. *Di is 0 Gy in the first cycle (i = 0) when the natural dose is measured. Preheat T and ITL T were determined experimentally (see text).

Stage	Treatment	Record
1	Administer regeneration dose D _i *	-
2	Preheat for 10 s at T°C	-
3	ITL at T°C for 500 s	ITL L _i
4	Give fixed test dose D_{t}	-
5	Preheat for 10 s at T°C	-
6	ITL at T°C for 500 s	ITL T _i
7	Repeat steps 1-6	-

Table 2: Details of the OSL age determinations including dose rate data, equivalent doses and resulting ages

[~]Derived using the 'central age model' (Galbraith et al., 1999) and reported with 1 standard error

Sample Details			Dosimetry Data			OSL Data		ITL Data	
Site	Lab code	Depth (m)	Cosmic dose (μGy a ⁻¹)	Water content (%)	Total dose rate (μGy a ⁻¹)	Average D _e ~ (Gy)	Age (ma)	Average D _e ~ (Gy)	Age (Ma)
Barrington Road	Shfd08191	3	170 ± 8	4.2	804 ± 66 ^{\$}	232 ± 6	0.29 ± 0.02	291 ± 15	0.36 ± 0.04
Makhulu Quarry	Shfd04287	1.6	174 ± 9	4.8	475 ± 20 [#]	248 ± 11*	>0.52#	806 ± 58*	>1.70

^{*} D_e values used to calculate a minimum age excludes aliquots which were saturated and as such represents a conservative estimation of minimum palaeodose

[#] published in Holmes et al (2007)

^{\$} determined using beta counting

Table 3: Bulk carbon and nitrogen contents, and δ^{13} C of the sampled lignite, along with *n*-alkane derived indices from the apolar solvent extraction.

Total organic carbon (%)	38.5
δ ¹³ C _{τος} (‰)	-26.6
Total Nitrogen (%)	0.8
TOC/TN	45
<i>n</i> -alkane Carbon Preference Index (CPI: C ₂₅ -C ₃₄)	4.57
n-alkane average chain length (C ₂₃ -C ₃₃)	27.9
P _{aq} (Ficken et al, 2000)	0.42

Table 4: Compound specific δ^{13} C data. Weighted mean *n*-alkane δ^{13} C is weighted by relative peak area.

Compound	Concentration (μg g ⁻¹)	δ ¹³ C (‰)		
C ₂₅ alkane	11.3	-25.6		
C ₂₆ alkane	2.57	-26.6		
C ₂₇ alkane	8.98	-25.2		
C ₂₈ alkane	2.09	-25.8		
C ₂₉ alkane	8.46	-28.1		
C ₃₁ alkane	13.9	-28.0		
C ₃₃ alkane	2.06	-27.3		
Weighted me	-27.0			
<i>Des-</i> A-Lupane	16.0	-24.4		
17β21β 30normoretane	1.99	-25.2		
17β 21β Homohopane	0.77	-20.6		

Table 5: Comparative table showing the Makhulu pollen data in relation previous reports of the Knysna Formation and Western Cape sites reported by Coetzee and Rogers (1982) and Cole and Roberts (1996) *Only pollen types listed in paper footnotes and/or referred to in Scott (1995) are shown here.

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		West Coast (Elandsfontein Formation: Miocene)				
Pollen type	Makhulu quarry	Coetzee et al. (1983)	Thiergart et al. (1963)	Helgren and Butzer (1977)*	Cole and Roberts (1996)	Coetzee and Rogers (1982)
Palm (Arecaceae)	Y (37%)	?		Υ		Υ
Cupressaceae						Υ
Podocarpus-sp	Y (3.7%)	Υ	Υ			Υ
Sapotaceae	Y (6.6%)					
Symphonia-type	Y (5.0%)					
Casuarinaceae		Υ				Υ
Cunonia?-type	Y (1.2%)					
Oleaceae			Υ			Υ
Ulmaceae						Υ
Poaceae	Y (4.6%)			Y (rare)	Υ	Υ
Euphorbiaceae	? (0.4%)		Y?			Υ
Asteraceae	Y (2.9%)				Υ	Υ
Anacardiaceae	Y (2.5%)		Υ			Υ
Rhus-type	Y (0.4%)	Υ	Υ		Υ	?
Rubiaceae	Y (0.4%)	?				Υ
Anthospermum- type	Y (0.4%)	?			Y	?
Myricaceae	Y (1.2%)					
Cliffortia-type	Y (0.4%)	Y			Υ	Υ
Cyperaceae	Y (6.7%)		Υ		Υ	
Ericaceae						
Proteaceae	? (0.4%)	Υ	Υ	Υ	Υ	Υ
Restionaceae		Υ	Υ		Υ	
Thymelaceae						?
Myrtaceae		Υ				