



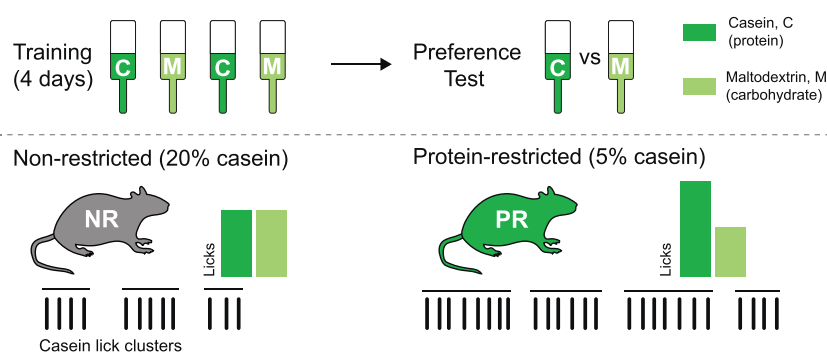
Restriction of dietary protein leads to conditioned protein preference and elevated palatability of protein-containing food in rats

Michelle Murphy, Kate Z. Peters, Bethany S. Denton, Kathryn A. Lee, Heramb Chadchankar, James E. McCutcheon*

Department of Neuroscience Psychology & Behaviour, University of Leicester, University Road, Leicester LE1 9HN, United Kingdom



GRAPHICAL ABSTRACT



ARTICLE INFO

Keywords:

Protein
Casein
Carbohydrate
Maltodextrin
Palatability
Microstructure

ABSTRACT

The mechanisms by which intake of dietary protein is regulated are poorly understood despite their potential involvement in determining food choice and appetite. In particular, it is unclear whether protein deficiency results in a specific appetite for protein and whether influences on diet are immediate or develop over time. To determine the effects of protein restriction on consumption, preference, and palatability for protein we assessed patterns of intake for casein (protein) and maltodextrin (carbohydrate) solutions in adult rats. To induce a state of protein restriction, rats were maintained on a low protein diet (5% casein) and compared to control rats on non-restricted diet (20% casein). Under these dietary conditions, relative to control rats, protein-restricted rats exhibited hyperphagia without weight gain. After two weeks, on alternate conditioning days, rats were given access to either isocaloric casein or maltodextrin solutions that were saccharin-sweetened and distinctly flavored whilst consumption and licking patterns were recorded. This allowed rats to learn about the post-ingestive nutritional consequences of the two different solutions. Subsequently, during a preference test when rats had access to both solutions, we found that protein-restricted rats exhibited a preference for casein over carbohydrate whereas non-restricted rats did not. Analysis of lick microstructure revealed that this preference was associated with an increase in cluster size and number, reflective of an increase in palatability. In conclusion, protein-restriction induced a conditioned preference for protein, relative to carbohydrate, and this was associated with increased palatability.

* Corresponding author.

E-mail address: jem64@le.ac.uk (J.E. McCutcheon).

1. Introduction

There is considerable evidence that of the three macronutrients dietary protein is most tightly regulated [1–3]. As such, when presented with diets that differ in macronutrient content, rats will adjust their consumption to ensure that protein intake meets a baseline level [4]. The mechanisms by which these adjustments occur are still not fully understood.

An important outstanding question is whether the drive for protein is immediate and innate or whether there is a role for learning using post-ingestive consequences [5,6]. Some evidence suggests that when protein-restricted a specific appetite for protein arises, similar to the appetite for sodium that arises under conditions of sodium depletion. Rats have been shown to rapidly increase their intake of a number of protein sources when protein-restricted in a manner that precludes using post-ingestive effects to guide their intake [7]. Further research suggested these rapid effects on protein appetite were driven by olfactory cues [8]. However, a large body of evidence indicates that adjustments to protein intake are slow, require experience with each food/diet, and likely involve post-ingestive feedback. For example, when allowed to select between diets that differ in protein content, it takes rats several days to adjust their intake appropriately [9]. This adaptation is more rapid in young rats, although still not immediate, presumably because protein requirements are elevated early in development and positive post-ingestive feedback is enhanced.

The majority of the above studies have assessed food intake and diet selection in home cage tests in which diets are given ad libitum. This arrangement does not allow precise monitoring of lick patterns over time. Sophisticated analysis of lick patterns, or lick microstructure, is a key method for assessing palatability of solutions in rodents [10]. As such, when individual licks are grouped into runs based on interlick intervals (termed bursts, clusters and bouts), increases in palatability are associated with longer runs of licking. Importantly, with respect to protein appetite, lick microstructure has not yet been investigated.

Learned shifts in the palatability of protein or protein-containing foods could contribute significantly to increased protein intake under protein-restriction. As a striking example, when rats are sodium-depleted normally aversive concentrations of sodium chloride become highly palatable [11]. Moreover, learning an association between conditioned flavors and intragastric infusions of glucose leads to an increase in palatability of the flavors paired with positive post-ingestive consequences [12,13]. However, increased intake is not always associated with shifts in palatability. For example, rats made deficient in a single essential amino acid increase their intake of the missing amino acid but this is not associated with an increase in palatability [14].

Here, we have used analysis of lick patterns to assess the effect of protein restriction on intake and palatability of isocaloric protein- and carbohydrate-containing solutions in adult rats. We find that protein-restricted rats, relative to controls, develop a learned preference for protein-containing solutions over carbohydrate and this is associated with an increase in relative palatability.

2. Materials and methods

2.1. Animals

Forty adult male Sprague-Dawley rats were used for experiments (Charles River; > 275 g at start of experiment). Twenty-four of these rats were used for the main behavioral experiment and a further sixteen contributed to the food intake data. Rats were group-housed (2–3 per cage) in IVCs with bedding materials as recommended by NC3R guidelines. Temperature was $21 \pm 2^\circ\text{C}$ and humidity was 40–50% with 12 h:12 h light/dark cycle (lights on at 07:00). Water was available ad libitum; chow containing different protein:carbohydrate ratio was available ad libitum (details below). All experiments were covered by the Animals [Scientific Procedures] Act (1986) and carried out

Table 1

Experimental diets used in study. List of ingredients (upper) and macronutrient breakdown (lower) in control diet (#D11051801; 20% casein) and protein-restricted diet (#D11092301; 5% casein).

Ingredient	D11051801 (control, 20% casein)	D11092301 (protein-restricted, 5%)	
	g/kg	g/kg	
Casein	200	50	
L-Cystine	3	0.75	
Corn starch	375.7	485	
Maltodextrin 10	125	150	
Sucrose	107.1	107.1	
Cellulose	50	50	
Soybean oil	25	25	
Lard	75	75	
Mineral mix S10022C	3.5	3.5	
Calcium carbonate	12.5	8.7	
Calcium phosphate, dibasic	0	5.3	
Potassium citrate	2.48	2.48	
Potassium phosphate, monobasic	6.86	6.86	
Sodium chloride	2.59	2.59	
Vitamin mix V10037	10	10	
Choline Bitartrate	2.5	2.5	
FD&C Yellow dye #5	0.05	0	
FD&C Red dye #40	0	0.05	

	g (%)	kcal (%)	g (%)	kcal (%)
Protein	18	18	5	4
Carbohydrate	62	60	76	74
Fat	10	22	10	22

under the appropriate license authority (Project License: 70/8069).

2.2. Diet manipulations

All rats were initially maintained on standard laboratory chow containing 20% dietary casein. To induce a state of protein restriction in half of the rats, standard chow was switched for one of two experimental diets based on modified AIN-93G that differed in protein:carbohydrate ratio (Table 1) but were isocaloric (4.1 kcal/g). Non-restricted diet (#D11051801, Research Diets, New Brunswick, NJ) contained 20% casein whereas protein-restricted diet (#D11092301, Research Diets) contained 5% casein. Body weight data were collected daily throughout the experiments. As rats were group-housed, food intake data were collected by cage and divided by the number of rats in the cage to give an average intake per animal. Conditioning experiments started 2 weeks following diet switch.

2.3. Behavioral testing

All testing took place within standard operant chambers (in cm: 30.5L, 24.1D, 21.0H; Med Associates, St. Albans City, VT) equipped with a house light and two bottles. Each bottle was connected to a contact lickometer calibrated to detect individual licks. Licks were recorded on a computer for all sessions as a measure of intake. All sessions lasted for one hour. For one to three days at the start of each experiment, rats were placed in the chambers with 0.2% sodium saccharin in both bottles to familiarize them with the apparatus. Following this, rats underwent a series of conditioning sessions and a preference test. In conditioning sessions, which occurred in a block of 4 days, only one bottle each day was available and was filled with either protein-containing solution (4% casein + 0.21% methionine + 0.2% sodium saccharin + 0.05% flavored Kool-Aid) or an isocaloric carbohydrate-containing solution (4% maltodextrin + 0.2% sodium saccharin

+ 0.05% flavored Kool-Aid) on alternate days. Methionine was added to the protein-based solution to make up for the relatively low levels of this amino acid present in casein [3]. Flavors (cherry vs. grape Kool-Aid) associated with each macronutrient and order of presentation (protein on days 1 and 3 vs. carbohydrate on days 1 and 3) were counter-balanced. In preference test sessions, both bottles and test solutions were available.

2.4. Analysis and statistical methods

Lick timestamp data from all experiments were analyzed in Python. All data files and custom scripts are available as supplemental files and are deposited on Github (<https://github.com/jaimemcc/murphy-2017>) and Mendeley Data (doi:<http://dx.doi.org/10.17632/wgd83v3ntb.1>). Lick microstructure was analyzed by using interlick intervals to divide licks into clusters [10]. Clusters were defined as runs of licks with no interlick intervals > 500 ms.

Body weight data were analyzed using two-way mixed ANOVA with diet as between-subjects factor and day as repeated measure. Food intake data were analyzed with cage as the statistical unit using an unpaired Student's *t*-test. Lick data for conditioning days, preference test, and measures of palatability were analyzed using two-way ANOVA with dietary group (non-restricted vs. protein-restricted) as between-subjects factor and solution (casein vs. maltodextrin) as within-subjects factor. On the preference test day, protein preference was calculated as licks for casein divided by total licks. Non-restricted vs. protein-restricted rats were compared using unpaired Student's *t*-test. For all analyses, α was set at 0.05 and all tests were two-tailed.

3. Results

3.1. Food intake and body weight data across low protein/high protein

First, we assessed whether maintenance on protein-restricted diets affected food intake and body weight of adult rats (Fig. 1). To date, much of the work on protein restriction has used younger rats when protein requirements are greater than in true adulthood. Here, we examined data from rats following the initial dietary manipulation but before conditioning sessions had started so that intake during these sessions did not confound our interpretations.

No difference in body weight was observed between the diet groups over the course of the experiment (Fig. 1A). As such, two-way ANOVA revealed a main effect of Day ($F(16,352) = 17.371$, $p < 0.001$) but no main effect of Diet ($F(1,22) = 0.115$, $p = 0.738$) and no Diet \times Day interaction ($F(16,352) = 0.574$, $p = 0.903$).

As rats were group-housed, we obtained food intake data by cage.

Food intake data from the eight cages of rats (three rats per cage) that participated in the main study are shown in Fig. 1B and visual inspection suggests a slight increase in intake (hyperphagia) in rats on protein-restricted diet. However, the small number of data points precludes statistical analysis. To address this, we combined this data set with food intake data from a pilot experiment in which an additional eight cages of rats were monitored (two rats per cage) and examined this extended data set (Fig. 1C). Statistical analysis of these data showed that protein-restricted rats did increase their intake of the low protein diet, relative to intake of non-restricted rats ($t(15) = 3.179$, $p = 0.007$). Thus, restriction of dietary protein resulted in hyperphagia without changes in body weight.

3.2. Protein restriction leads to development of preference for protein-containing solutions

Next, we asked whether rats would display a greater preference for protein-containing solutions over carbohydrate-containing solutions when they were protein-restricted. Our experiment was divided into conditioning days when only one solution was available – casein or maltodextrin – followed by a preference test day when both solutions were available. First, we analyzed data from conditioning days for both casein and maltodextrin to look for effects of diet or conditioning day (Fig. 2).

Analysis of data from casein conditioning days (Fig. 2A and B) revealed a significant interaction between Diet and Day ($F(1,22) = 7.222$, $p = 0.014$) with no main effects of Diet ($F(1,22) = 1.19$, $p = 0.287$) or Day ($F(1,22) = 0.633$, $p = 0.435$). Further analysis of each day separately revealed that on Day 1 there was a trend towards protein-restricted rats drinking more casein than non-restricted rats ($t(11) = 1.97$, $p = 0.061$). Interestingly, visual inspection of the time course data (Fig. 2A) suggests that on Day 1 protein-restricted rats showed a different pattern of casein consumption than non-restricted rats. As such, both groups of rats drank a similar amount in the first twenty minutes of the session but in the final forty minutes, casein consumption appeared to increase in protein-restricted rats, relative to non-restricted rats. Casein consumption on the second conditioning day did not differ between protein-restricted and non-restricted rats ($t(11) = 0.160$, $p = 0.874$).

Analysis of consumption during maltodextrin conditioning sessions (Fig. 2C and D) showed that protein-restricted rats drank more maltodextrin on both conditioning days than non-restricted rats. As such, there was a main effect of Diet ($F(1,22) = 4.825$, $p = 0.039$) with no main effect of Day ($F(1,22) = 0.222$, $p = 0.642$) and no significant interaction ($F(1,22) = 2.343$, $p = 0.140$).

Finally, when we analyzed total consumption of casein and

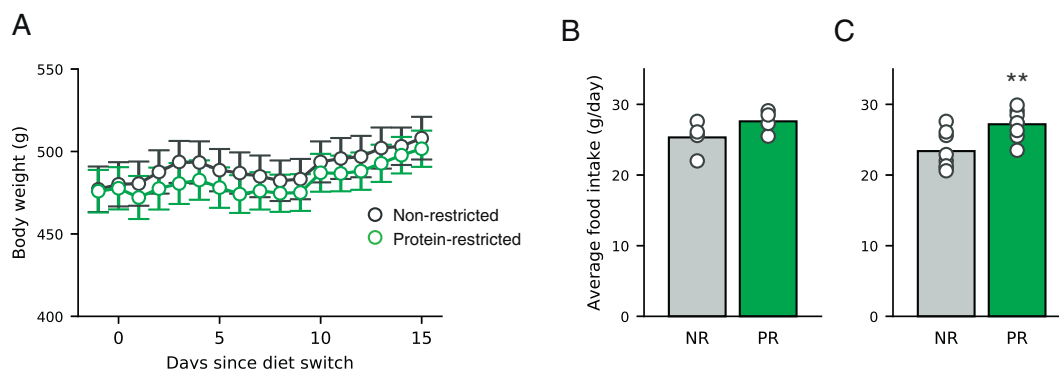


Fig. 1. Protein-restricted adult rats increase food intake without changes in body weight. (A) Body weight gradually increases over the course of the experiment in non-restricted (NR; black) and protein-restricted (PR; green) rats with no difference between groups. Data are mean \pm SEM. (B) Food intake is greater in protein-restricted rats, relative to non-restricted rats. Intake is shown as grams per day per rat calculated by dividing total daily intake by number of rats in a cage. Bars show mean and data from individual cages are shown as circles. (C) Same data as in (B) supplemented with food intake data from a pilot experiment using rats of comparable age and weight. $**p < 0.01$ vs. non-restricted rats. [figure = 2 columns]. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

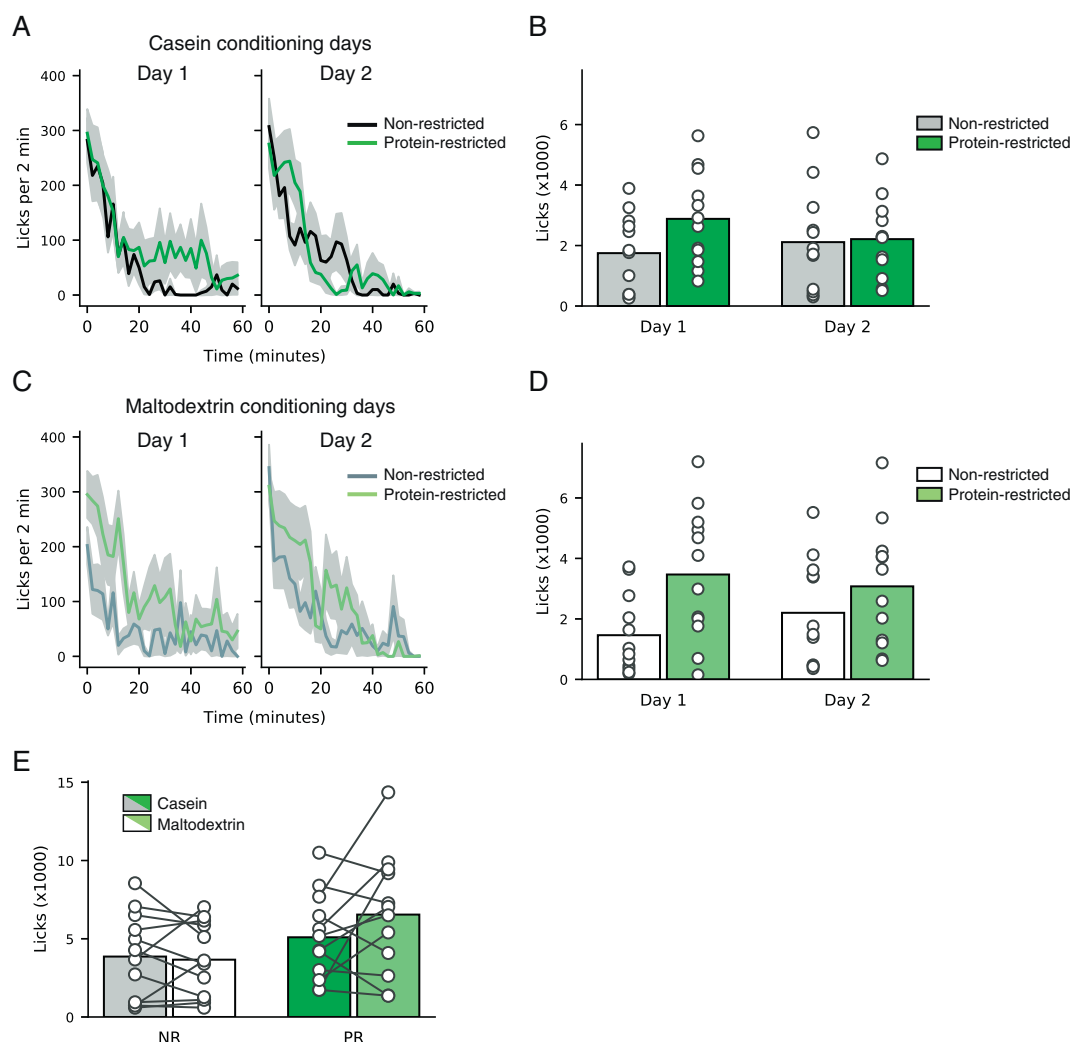


Fig. 2. Protein-restricted and non-restricted rats drink similar amounts during conditioning sessions. Rats were given access to distinctly-flavored casein (4%; 2×1 h sessions) or maltodextrin (4%; 2×1 h sessions) over four days. (A) Time course of licking for casein on conditioning days 1 and 2 during each 1 h conditioning session. Lick rates for non-restricted (black line) and protein-restricted (dark green) rats are shown. Lines are mean and shaded area is SEM. (B) Licks of casein on conditioning days 1 and 2 in non-restricted and protein-restricted rats. (C) Time course of licking for maltodextrin on conditioning days 1 and 2 during each 1 h conditioning session. Lick rates for non-restricted (dark grey) and protein-restricted animals (light green) are shown. Lines are mean and shaded area is SEM. (D) Licks of maltodextrin on conditioning days 1 and 2 in non-restricted and restricted rats. (E) Total licks over both sessions for non-restricted (NR, grey and white bars) and protein-restricted (PR, green bars). Dark bars show casein licks and light/white bars show maltodextrin licks. For all panels, bars represent mean and circles are data from individual rats. [figure = 2 columns]. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

maltodextrin across conditioning sessions (Fig. 2E) we found no significant differences between protein-restricted and non-restricted rats although there was a trend for protein-restricted rats to drink more of both solutions than non-restricted rats. As such, two-way mixed ANOVA revealed a trend towards a main effect of Diet ($F(1,22) = 3.609$, $p = 0.071$) but no main effect of Solution ($F(1,22) = 1.203$, $p = 0.285$) and no interaction between Diet and Solution ($F(1,22) = 2.087$, $p = 0.163$). In summary, there were subtle differences in consumption between diet groups when only one solution was available there was no clear difference in preference for protein over carbohydrate on conditioning days.

Following these four conditioning sessions, rats were given access to both solutions during the same session (Fig. 3). In this session, protein-restricted rats drank more casein than maltodextrin and this elevated intake appeared to occur in the first twenty minutes of the session (Fig. 3A). Furthermore, protein-restricted rats showed a significant preference for casein over maltodextrin whereas non-restricted rats did not (Fig. 3B & 3C). As such, two-way ANOVA revealed that there was a main effect of Solution ($F(1,22) = 7.466$, $p = 0.012$) and an

interaction between Solution and Diet ($F(1,22) = 11.677$, $p = 0.002$).

Subsequent analysis of each diet group individually showed that protein-restricted rats licked more for casein than maltodextrin ($t(11) = 4.630$, $p < 0.001$) but non-restricted rats did not ($t(11) = 0.458$, $p = 0.656$). In addition, we calculated a casein preference score by dividing casein licks by total licks (Fig. 3D) and found that protein-restricted rats showed a greater protein preference, relative to non-restricted rats ($t(21) = 2.660$, $p = 0.015$).

3.3. Palatability of protein-containing solutions is increased by protein-restriction

Finally, we used analysis of lick microstructure [10] to examine whether the palatability of protein-containing solutions was affected by the state of protein restriction. Lick patterns were divided into clusters, separated by interlick intervals > 500 ms. An increased number of licks per cluster is generally thought to reflect increased palatability. We found that the state of protein restriction influenced palatability of casein, relative to maltodextrin (Fig. 4). As such, two-way ANOVA

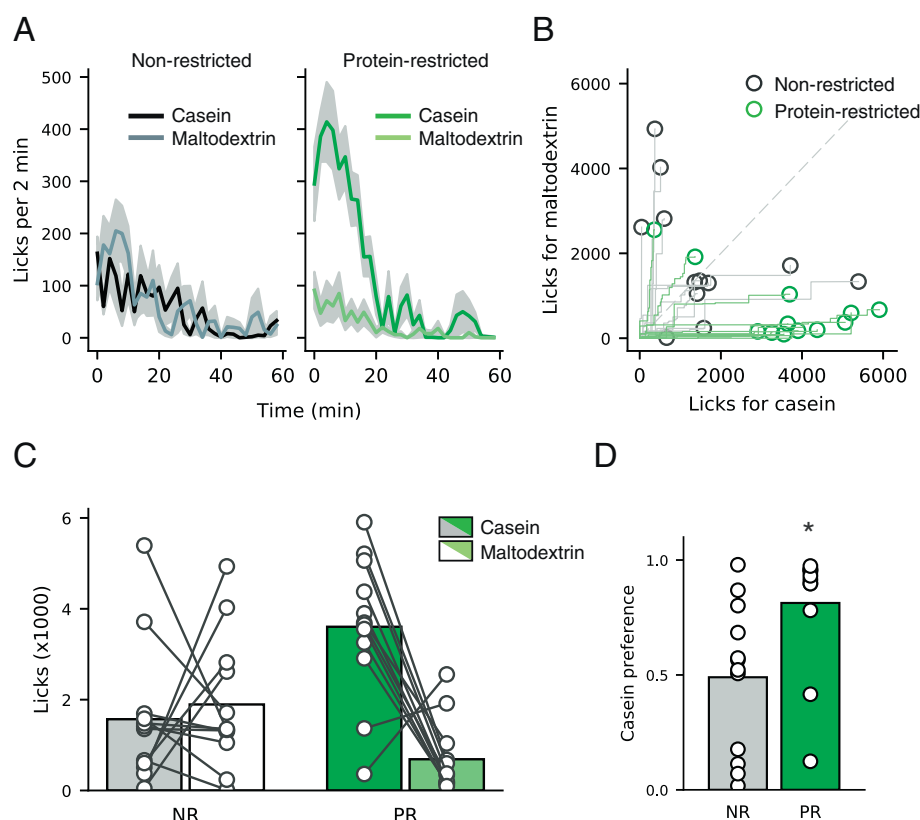


Fig. 3. Protein-restricted rats show preference for protein over carbohydrate. After conditioning sessions rats were given access to both casein and maltodextrin solutions within the same session. (A) Time course of licking for casein and maltodextrin during the 1 h preference session. Similar rates of licking are seen for both casein (black line) and maltodextrin (grey line) in non-restricted rats whereas in protein-restricted rats elevated licking is observed to casein (dark green) vs. maltodextrin (light green). This licking predominantly occurs in the first twenty minutes of the session. Lines are mean and shaded area shows SEM. (B) Cumulative licks for casein vs. maltodextrin are shown for individual rats that were non-restricted (grey lines and black circles) or protein-restricted (green lines and circles). Consecutive licks are plotted with casein licks advancing along the x-axis and maltodextrin licks along the y-axis. Dashed grey line at unity represents absence of preference for either solution whereas markers to the right represent casein preference and markers to the left maltodextrin preference. The majority of protein-restricted rats lie to the right of this line indicating protein preference whereas non-restricted rats are evenly distributed. (C) Licks of casein vs. maltodextrin during preference session. Conventions are identical to Fig. 2. (D) Casein preference calculated as casein licks divided by total licks. Protein-restricted rats (green bar) show an increased preference for casein, relative to non-restricted rats (grey bar). Bars are mean and circles are data from individual rats. * $p < 0.05$ vs. non-restricted rats. [figure = 1.5 columns]. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

revealed a significant interaction between Solution and Diet ($F(1,22) = 7.099$, $p = 0.014$).

Further analysis of each diet group separately showed that casein and maltodextrin had similar palatability in non-restricted rats ($t(11) = 0.761$, $p = 0.463$) but the palatability of casein was elevated relative to maltodextrin in protein-restricted rats ($t(11) = 2.688$, $p = 0.021$).

In addition, the number of clusters was also influenced by the state of protein-restriction as two way ANOVA revealed a main effect of Solution ($F(1,22) = 5.677$, $p = 0.026$) and an interaction between Solution and Diet ($F(1,22) = 7.119$, $p = 0.014$). Analysis of each diet group separately showed that in non-restricted rats there were the same number of clusters for both casein and maltodextrin ($t(11) = 0.203$, $p = 0.843$) whereas protein-restricted rats had an increased number of clusters for casein, relative to maltodextrin ($t(11) = 3.550$, $p = 0.005$).

4. Discussion

Here, we examined the effect of protein restriction on development of preference and palatability of protein- vs. carbohydrate-containing solutions. We found that maintenance on a protein-restricted diet resulted in rats developing a preference for protein vs. carbohydrate when given a choice between the two. Moreover, the increase in protein intake was associated with an increase in palatability of the protein-containing solution, relative to the carbohydrate-containing solution.

We monitored food intake and body weight for the two weeks following the change to a protein-restricted diet but before beginning behavioral sessions. Previous studies have found that rats on diets that are moderately low in protein show hyperphagia without weight gain [9,15,16]. In support of these studies, we found that protein-restricted rats increased food intake, relative to controls, without changing their body weight. It is of note, however, that the slight increase in food intake we observe is still far below what is needed to match the protein intake of control, non-restricted rats. In our studies, we used a low protein diet that contained 5% protein whereas other studies using rats

have found effects on behavioral and metabolic parameters using diets containing 10% protein [15]. Our choice of 5% was based on pilot experiments, in which we found no effects of 10% protein diet on food intake or conditioned preferences in adult rats (data not shown). The likely explanation for this variation in effective dietary manipulations is different protein requirements during development. Many studies have used late adolescent or young adult rats rather than mature animals and differences in the effects of low protein diets across age and development are well documented [9,17].

In conditioning sessions, over four days rats were given one type of solution - containing either protein or carbohydrate - and lick patterns were monitored. Although in general, rats from both dietary conditions drank similar amounts of casein and maltodextrin during these sessions subtle differences between the groups were apparent. For example, on the first casein conditioning day there was a suggestion that protein-restricted rats drank more casein than non-restricted rats especially during the late part of the session. This late consumption of casein could reflect an appetitive post-ingestive effect of casein. In addition, analysis of maltodextrin consumption or total consumption suggested that in other circumstances protein-restricted rats drank slightly more of both solutions than control rats. This may reflect a moderate form of hyperphagia, similar to home cage intake reported above. Intriguingly, in these conditioning sessions, when only one solution was available, consumption was increased for the carbohydrate-containing solution meaning that protein-restriction may also generate a hyperphagic response that disregards the macronutrient content of the food on offer.

One potential explanation for this lack of preference in conditioning sessions when only one bottle is available may be that rats are consuming close to their maximal intake during these hour-long sessions and thus satiety mechanisms are engaged that prevent further consumption. Another possibility is that when both solutions are presented together the comparison generates a negative contrast effect whereby the value of the maltodextrin solution is reduced, relative to the casein solution [18].

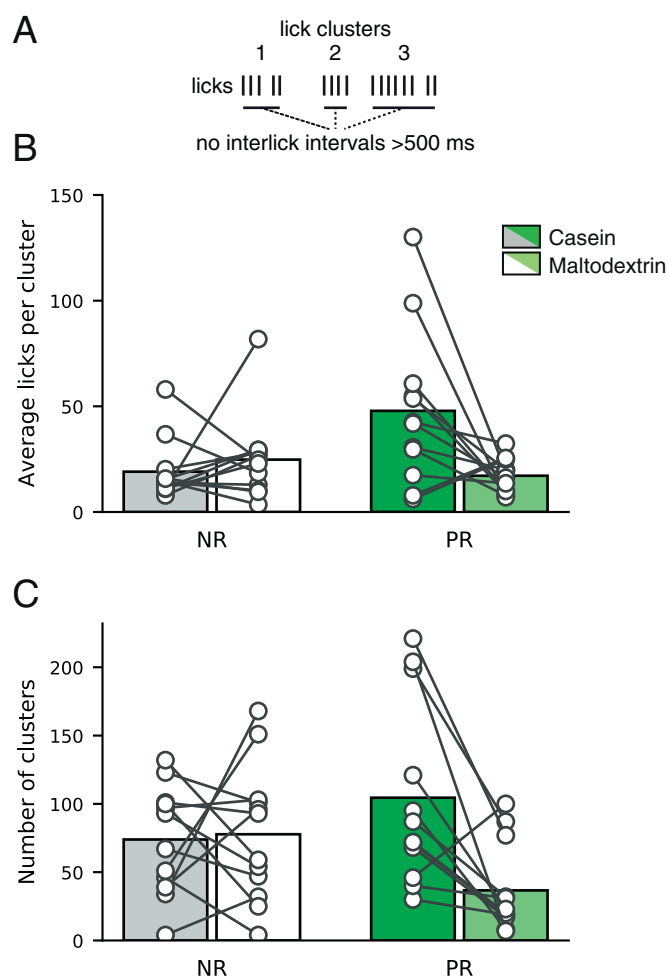


Fig. 4. Palatability of protein is enhanced in protein-restricted rats relative to carbohydrate. (A) Schematic showing criteria for defining lick clusters. Licks were grouped into clusters based on having interlick intervals < 500 ms. (B) Average licks per clusters in preference test are shown for casein (dark bars) and maltodextrin (light/white bars) for non-restricted (NR; grey/white) and protein-restricted rats (PR; green). Protein-restricted rats show elevated licks per cluster for casein, relative to maltodextrin. Bars are mean and circles are individual rats. (C) Number of clusters is shown for casein and maltodextrin in non-restricted and protein-restricted rats. Conventions are identical to (B). [figure = 1 column]. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

In the preference test, when rats were given access to both solutions, we found a strong preference towards the protein-containing solution in protein-restricted rats. This preference was not present in control rats. This finding corroborates other work showing that protein-restricted rats can direct their behavior to increase protein intake. In addition, we have extended these previous studies by analyzing the precise temporal patterns of licking to assess how lick macrostructure and microstructure are affected by protein restriction. By analyzing lick microstructure during the preference test, we found that palatability of the protein-containing solutions increased in protein-restricted rats indicating that this might be a mechanism that drives increased intake of protein-containing foods. This situation parallels studies that examined palatability after flavor-nutrient conditioning. When flavored saccharin is paired with intragastric glucose infusions, palatability of the paired flavor is elevated [12]. Our studies used a similar paradigm in which solutions were sweetened with saccharin and distinctly-flavored with Kool-Aid, as is common in studies of flavor-nutrient conditioning [19]. Thus, increased palatability (flavor evaluation) might be a mechanism that drives increased intake by promoting more meals and longer meals.

The presentation of macronutrients in combination with saccharin

and flavoring means that we do not know whether the changes in palatability that we observe reflect a change in palatability of individual components of the solution or the combination. When rats are made sodium-deficient, the nutrient itself, sodium, immediately becomes more palatable in an experience-independent manner [11]. This shift is profound as it applies to high concentrations of sodium, which are normally evaluated as aversive in sodium-replete animals. Moreover, sodium-evoked dopamine signals and appetitive behavioral responses to sodium-associated cues also emerge with no experience of sodium in a depleted state [20,21]. Literature suggests that appetite for protein may differ from sodium appetite. For example, when rats are maintained on a diet deficient in a single essential amino acid (lysine), they develop compensatory responses, which increase their intake of lysine, but these responses take ~30 min to emerge and longer if they are required to discriminate between two different amino acids [14]. Interestingly, in this study no evidence of an increase in palatability, assessed by bout size, was observed. In addition, this delayed rather than immediate time course for the development of protein-directed responses suggests that learning about the post-ingestive consequences of protein ingestion is essential in a manner that is fundamentally different from sodium appetite. Our data, although not conclusive, support this idea as casein consumption on the first day of experience trends towards being different between protein-restricted and non-restricted rats but consumption of the two dietary groups only begins to diverge late (20–30 min) into the session.

A further point of consideration is our use of casein as the sole source of protein in these studies. We have used casein as it is the most commonly studied protein source in experiments of this kind and it comprises the major protein source in most standard laboratory diets for rodents. It has a relatively well-balanced profile of amino acids with the exception of cysteine/methionine, which is why we added methionine to all casein-containing solutions [3]. It will be of great interest to determine in future studies whether other protein sources drive behavioral preference to a similar level as other sources with different sensory and absorptive properties may differ. Indeed, previous studies indicated variable responses to different protein sources including soybean, gluten, and gelatin, that may be driven by their olfactory properties [7,8].

Protein is a vital source of essential amino acids which are crucial precursors of neurotransmitters and are involved in almost all essential bodily processes. The importance of individual amino acids and their roles in regulating protein appetite and satiation is an active area of study. The avoidance of toxic levels of amino acids may underlie the aversive nature of very high protein diets – and their subsequent anorectic effects on consumption [22]. Conversely the maintenance of a minimum or optimal level of particular amino acids may contribute to the effects of protein restriction on increasing intake [6].

Specific amino acids can be detected by orosensory cues such as taste and by specific receptors, for example glutamate directly interacts with its own metabotropic receptors [23]. They are also sensed at the level of the duodenum and intestine and affect gut hormones [24]. Rats given diets deficient in single amino acids are capable of distinguishing and selecting for that specific amino acid in choice tests [25,26]. The existence of a central amino acid sensing mechanism has been suggested involving the anterior piriform cortex [22,27] and there appears to be a role for both central and peripheral mediators. Whether levels of individual amino acids contribute to the changes in consumption we have observed during dietary protein restriction remains to be elucidated.

One of the most thought-provoking theories developed to explain the obesity crisis is the protein leveraging hypothesis [28,29]. This theory posits that a steady decrease in the proportion of protein in Western diets occurring over the last few decades has resulted in carbohydrate and fat being overconsumed. The relatively minor role of protein in overall energy intake (generally < 20%) produces this leveraging ability and means that compensating for even small changes

in protein can lead to significant overconsumption of energy from fat and carbohydrate. An important assumption of this hypothesis is that deficiencies in specific nutrients influence our feeding behavior by triggering consumption but that this consumption is indiscriminate and not well-targeted to replenish the nutrient in deficit. Contrary to this assumption, our data suggest that, at least in rats, protein-restriction does recruit mechanisms that enable rats to guide their behavior towards consumption of protein-rich food. However, our studies are far from modelling the human situation and there are numerous important discrepancies to be addressed. First, the level of protein restriction is likely more severe in our protocol than that which most humans in the developed world encounter. Second, the choice of food provided in our studies was limited (protein vs carbohydrate with similar sweetness but distinct flavor) and did not include foods that contained a mixture of macronutrients. Third, the pattern of experience (each solution separately on alternate days) was designed to maximize the ability of rats to discriminate post-ingestive effects and learn about the nutritional value of each solution. In the human situation, where foods contain mixtures of macronutrients and other flavorings, fine discrimination of nutritional consequences of ingestion is likely far more difficult. Moreover, numerous other factors influence our intake such as social setting, cultural norms and access, which may bias us against choosing food stuffs based solely on nutritional outcome. Future studies will attempt to address the ability of rats to develop protein preferences in more challenging situations that better model the human context.

Acknowledgements

Funding: This work was supported by the Biotechnology and Biological Sciences Research Council [grant # BB/M007391/1]; and the European Commission [grant # GA 631404].

Data statement

All raw data files will be published with this manuscript alongside the Python scripts used to perform analysis. These are deposited on Github (<https://github.com/jaimemcc/murphy-2017>) and Mendeley Data (doi:<http://dx.doi.org/10.17632/wgd83v3ntb.1>).

References

- [1] H.-R. Berthoud, H. Münzberg, B.K. Richards, C.D. Morrison, Neural and metabolic regulation of macronutrient intake and selection, *Proc. Nutr. Soc.* 71 (2012) 390–400, <http://dx.doi.org/10.1017/S0029665112000559>.
- [2] C.P. Richter, Total self-regulatory functions in animals and human beings, *Harvey Lect.* 38 (1943) 63–103.
- [3] C.L. Theall, J.J. Wurtman, R.J. Wurtman, Self-selection and regulation of protein: carbohydrate ratio in foods adult rats eat, *J. Nutr.* 114 (1984) 711–718.
- [4] J.C. Peters, A.E. Harper, Adaptation of rats to diets containing different levels of protein: effects on food intake, plasma and brain amino acid concentrations and brain neurotransmitter metabolism, *J. Nutr.* 115 (1985) 382–398.
- [5] B.G. Galef Jr., Is there a specific appetite for protein? *Neural Metab. Control Macromol. Intake.* (2000) 18–28.
- [6] C.D. Morrison, T. Laeger, Protein-dependent regulation of feeding and metabolism, *Trends Endocrinol. Metab.* 26 (2015) 256–262, <http://dx.doi.org/10.1016/j.tem.2015.02.008>.
- [7] J.A. Deutsch, B.O. Moore, S.C. Heinrichs, Unlearned specific appetite for protein, *Physiol. Behav.* 46 (1989) 619–624, [http://dx.doi.org/10.1016/0031-9384\(89\)90341-7](http://dx.doi.org/10.1016/0031-9384(89)90341-7).
- [8] S.C. Heinrichs, J.A. Deutsch, B.O. Moore, Olfactory self-selection of protein-containing foods, *Physiol. Behav.* 47 (1990) 409–413, [http://dx.doi.org/10.1016/0031-9384\(90\)90101-9](http://dx.doi.org/10.1016/0031-9384(90)90101-9).
- [9] B.D. White, M.H. Porter, R.J. Martin, Effects of age on the feeding response to moderately low dietary protein in rats, *Physiol. Behav.* 68 (2000) 673–681, [http://dx.doi.org/10.1016/S0031-9384\(99\)00229-2](http://dx.doi.org/10.1016/S0031-9384(99)00229-2).
- [10] J.D. Davis, G.P. Smith, Analysis of the Microstructure of the Rhythmic Tongue Movements of Rats Ingesting Maltose and Sucrose Solutions, 106 (1992), pp. 217–228.
- [11] K.C. Berridge, F.W. Flynn, J. Schulkin, H.J. Grill, Sodium depletion enhances salt palatability in rats, *Behav. Neurosci.* 98 (1984) 652–660.
- [12] K.P. Myers, A. Sclafani, Conditioned enhancement of flavor evaluation reinforced by intragastric glucose: I. Intake acceptance and preference analysis, *Physiol. Behav.* 74 (2001) 481–493, [http://dx.doi.org/10.1016/S0031-9384\(01\)00595-9](http://dx.doi.org/10.1016/S0031-9384(01)00595-9).
- [13] K.P. Myers, A. Sclafani, Conditioned enhancement of flavor evaluation reinforced by intragastric glucose. II. Taste reactivity analysis, *Physiol. Behav.* 74 (2001) 495–505, [http://dx.doi.org/10.1016/S0031-9384\(01\)00595-9](http://dx.doi.org/10.1016/S0031-9384(01)00595-9).
- [14] S. Markison, B.L. Thompson, J.C. Smith, A.C. Spector, Time course and pattern of compensatory ingestive behavioral adjustments to lysine deficiency in rats, *J. Nutr.* 130 (2000) 1320–1328.
- [15] T. Laeger, T.M. Henagan, D.C. Albarado, L.M. Redman, G.A. Bray, R.C. Noland, et al., FGF21 is an endocrine signal of protein restriction, *J. Clin. Invest.* 124 (2014) 3913–3922, <http://dx.doi.org/10.1172/JCI77508>.
- [16] C.D. Morrison, X. Xi, C.L. White, J. Ye, R.J. Martin, Amino acids inhibit AgRP gene expression via an mTOR-dependent mechanism, *Am. J. Physiol. Endocrinol. Metab.* 293 (2007) E165–71, <http://dx.doi.org/10.1152/ajpendo.00675.2006>.
- [17] P.D. Leathwood, D.V. Ashley, Strategies of protein selection by weanling and adult rats, *Appetite* 4 (1983) 97–112.
- [18] C. Mitchell, C. Flaherty, Temporal Dynamics of Corticosterone Elevation in Successive Negative Contrast, 64 (1998), pp. 287–292.
- [19] A. Sclafani, K. Touzani, R.J. Bodnar, Dopamine and learned food preferences, *Physiol. Behav.* 104 (2011) 64–68, <http://dx.doi.org/10.1016/j.physbeh.2011.04.039>.
- [20] J.J. Cone, S.M. Fortin, J.A. McHenry, G.D. Stuber, J.E. McCutcheon, M.F. Roitman, Physiological state gates acquisition and expression of mesolimbic reward prediction signals, *Proc. Natl. Acad. Sci. U. S. A.* 113 (2016) 1943–1948, <http://dx.doi.org/10.1073/pnas.1519643113>.
- [21] M.J.F. Robinson, K.C. Berridge, Instant transformation of learned repulsion into motivational “wanting”, *Curr. Biol.* 23 (2013) 282–289, <http://dx.doi.org/10.1016/j.cub.2013.01.016>.
- [22] A. Blais, L.J. Magrum, T.J. Koehnle, J.W. Sharp, D. Tome, D.W. Gietzen, Biochemical and Molecular Actions of Nutrients Threonine Deprivation Rapidly Activates the System A Amino Acid Transporter in Primary Cultures of Rat Neurons from the Essential Amino Acid Sensor in the Anterior Piriform Cortex 1, 2, 2 (2003), pp. 2156–2164.
- [23] G. Fromentin, N. Darcel, C. Chaumontet, P.C. Even, D. Tome, C. Gaudichon, Control of food intake by dietary amino acids and proteins: molecular and cellular aspects, *Mol. Nutr. Amin. Acids Proteins* (2016) 221–232.
- [24] R. Rasoamanana, N. Darcel, G. Fromentin, D. Tome, 70th Anniversary: Body weight regulation – food, gut and Brain Signalling Symposium I: Food – gut Interactions Nutrient Sensing and Signalling by the gut Proceedings of the Nutrition Society, (2017), pp. 446–455, <http://dx.doi.org/10.1017/S0029665112000110>.
- [25] B.J. Hrupka, Y.M. Lin, D.W. Gietzen, Q.R. Rogers, Nutrient Requirements and Interactions Small Changes in Essential Amino Acid Concentrations Alter Diet Selection in Amino Acid – Deficient Rats 1, 2, (1997), pp. 777–784.
- [26] K. Torii, A. Nijijima, Effect of Lysine on Afferent Activity of the Hepatic Branch of the Vagus Nerve in Normal and L-lysine-Deficient Rats, 72 (2001), pp. 685–690.
- [27] C. Amino, A. Deficiency, D.W. Gietzen, L.J. Magrum, Symposium: Leucine as a Nutritional Signal Molecular Mechanisms in the Brain Involved in the Anorexia of Branched, (2001), pp. 851–855.
- [28] S.J. Simpson, D. Raubenheimer, Obesity: the protein leverage hypothesis, *Obes. Rev.* 6 (2005) 133–142, <http://dx.doi.org/10.1111/j.1467-789X.2005.00178.x>.
- [29] S.J. Simpson, R. Batley, D. Raubenheimer, Geometric analysis of macronutrient intake in humans: the power of protein? *Appetite* 41 (2003) 123–140, [http://dx.doi.org/10.1016/S0195-6663\(03\)00049-7](http://dx.doi.org/10.1016/S0195-6663(03)00049-7).